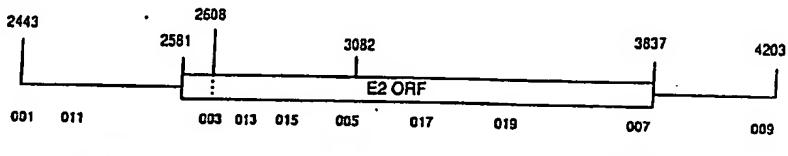




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(54) Title: ANTISENSE OLIGONUCLEOTIDE INHIBITION OF PAPILLOMAVIRUS



Oligo	5'	3'	Comments
LC001.ABI	AGGTTTGCACCCGACTATGCAAGTACAAT		mRNA cap region
LC002.ABI	TATGCAAGTACAAT		mRNA cap region
LC003.ABI	CGTTCGCATGCTCTCCATCCCTCTTCACT		Initiation of translation
LC004.ABI	GCATGCTGTCTCCAT		Initiation of translation
LC005.ABI	AAATCGTCAGCACGGGCCATGGTGCAGT		Transrepressor start
LC006.ABI	AGCACCGGGCATGGT		Transrepressor start
LC007.ABI	CAATGCGACTGATCAGAAGTCAAAGCTGGC		translational termination
LC008.ABI	GCAGCTGATCAGAAGT		translational termination
LC009.ABI	ATTGCTGCAGCTTAAACCATATAAAATCTG		3' untranslated region
LC010.ABI	CTTAAACCATATAAA		3' untranslated region
LC011.ABI	AAAAAAAGATTCCCAAJCTGCATCAGTAAT		5' untranslated region
LC012.ABI	AAGATTCCAATCTG		5' untranslated region
LC013.ABI	CAGTGTCCCTAGGACAGTCACCCCTTTTC		5' coding region
LC014.ABI	GGACAGTCACCCCTT		5' coding region
LC015.ABI	TGTACAAATTGCTGTAGACAGTGTACAGT		mid coding region
LC016.ABI	GCTGTAGACAGTGT		mid coding region
LC017.ABI	GTGCGAGGCGAGGACCGTCCCGTACCCAACC		3' coding region
LC018.ABI	GGACCGTCCCCGTAC		3' coding region
LC019.ABI	TTAACAGGTGGAATCCATCATGGTGGTG		5' coding region
LC020.ABI	GGATCCATCTTGG		5' coding region

(57) Abstract

Oligonucleotides are provided which are capable of antisense interaction with messenger RNA of papillomavirus. Such oligonucleotides or oligonucleotide analogs can be used for diagnostics and therapeutics as well as for research purposes. In accordance with preferred embodiments of this invention, oligonucleotides are provided which are hybridizable with selected messenger RNA from a human papillomavirus. The oligonucleotide is able to inhibit the function of the RNA, and accordingly is useful for therapy for infections by such papillomavirus. In accordance with a preferred embodiment, portions of the papillomavirus are targeted for antisense attack. Thus, oligonucleotides are preferably provided which hybridize with the E2, E6 and E7 messenger RNAs.

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**ANTISENSE OLIGONUCLEOTIDE INHIBITION
OF PAPILLOMAVIRUS**

FIELD OF THE INVENTION

This invention relates to the inhibition of papillomavirus and the diagnosis and treatment of infections in animals caused by papillomavirus. This invention is also directed to the detection and quantitation of papillomavirus in samples suspected of containing it. Additionally, this invention is directed to oligonucleotides and oligonucleotide analogs which interfere with or modulate the function of messenger RNA from papillomavirus. Such interference can be employed as a means of diagnosis and treatment of papillomavirus infections. It can also form the basis for research reagents and for kits both for research and for diagnosis.

BACKGROUND OF THE INVENTION

The papillomaviruses (PV) are widespread in nature and are generally associated with benign epithelial and fibroepithelial lesions commonly referred to as warts. They have been detected in and isolated from a variety of higher vertebrates including human, cattle, rabbits, deer and several avian species. Although these viruses are generally associated with benign lesions, a specific subset of the viruses have been associated with lesions that may progress

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to carcinomas. The implication that these viruses may play an etiologic role in the development of some human cancers follows from numerous studies that have shown the presence of transcriptionally active human papillomavirus (HPV) deoxyribo-
5 nucleic acids in a high percentage of certain cancerous lesions. Zur Hausen, H. and Schneider, A. 1987. In: The Papovaviridae, vol. 2, edited by N. P. Salzman and P. M. Howley, pp. 245-264. Plenum Press, New York.

In man, human papillomaviruses cause a variety of
10 disease including common warts of the hands and feet, laryngeal warts and genital warts. More than 57 types of HPV have been identified so far. Each HPV type has a preferred anatomical site of infection; each virus can generally be associated with a specific lesion. Genital warts, also
15 referred to as venereal warts and condylomata acuminata, are one of the most serious manifestations of PV infection. As reported by the Center for Disease Control, the sexual mode of transmission of genital warts is well established and the incidence of genital warts is on the increase. The
20 seriousness of genital warts is underlined by the recent discovery that HPV DNA can be found in all grades of cervical intraepithelial neoplasia (CIN I-III) and that a specific subset of HPV types can be found in carcinoma *in situ* of the cervix. Consequently, women with genital warts, containing
25 specific HPV types are now considered at high risk for the development of cervical cancer. Current treatments for genital warts are inadequate.

There is a great, but as yet unfulfilled, desire to provide compositions of matter which can interfere with
30 papillomavirus. It is similarly desired to achieve methods of therapeutics and diagnostics for papillomavirus infections in animals. Additionally, improved kits and research reagents for use in the study of papillomavirus are needed.

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OBJECTS OF THE INVENTION

It is an object of this invention to provide oligonucleotides which are capable of hybridizing with messenger RNA of papillomavirus to inhibit the function of the 5 messenger RNA.

It is a further object to provide oligonucleotides which can modulate the functional expression of papillomavirus DNA through antisense interaction with messenger RNA of the virus.

10 Yet another object of this invention is to provide methods of diagnostics and therapeutics for papillomavirus in animals.

Methods, materials and kits for detecting the presence or absence of papillomavirus in a sample suspected 15 of containing it are further objects of the invention.

Novel oligonucleotides are other objects of the invention.

These and other objects will become apparent to persons of ordinary skill in the art from a review of the 20 instant specification and appended claims.

BRIEF DESCRIPTION OF THE DRAWINGS

Figures 1 a, b, c and d, are schematic maps of the genetic organization of several PV genomes.

Figure 2 is a partial genetic mapping of a bovine 25 papillomavirus, BPV-1, genome showing open reading frames, ORFs, and messenger RNAs transcribed from the genome.

Figure 3 is a nucleotide sequence of the BPV-1 E2 transactivator gene mRNA showing nucleotides 2443 through 4203.

30 Figure 4 is a nucleotide sequence of the BPV-1 E2 transactivator gene mRNA showing the domain having nucleotides 2443 through 3080.

Figure 5 is a nucleotide sequence of the 5' common untranslated region of BPV-1 coding for early messenger RNAs 35 showing the domain having nucleotides 89 through 304.

Figure 6 is the nucleotide sequences of antisense

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oligonucleotides made in accordance with the teachings of the invention and the relative position of the oligonucleotides on the E2 mRNA. The oligonucleotide identifier, sequence and functional role are depicted.

5 Figure 7 is a graphical depiction of the effects of antisense oligonucleotides made in accordance with the teachings of the invention on E2 expression. Oligonucleotides targeted to the mRNA CAP region (I1751) and the translation codon for the E2 transactivator (I1753) are shown to reduce
10 E2 transactivation at micromolar concentrations.

Figure 8 is a graphical depiction of the dose response of antisense oligonucleotides made in accordance with the teachings of the invention. These dose response curves show that an antisense oligonucleotide, I1753, which is
15 complementary to the E2 transactivation messenger RNA in the region including the translation initiation codon has an 50% inhibitory concentration (IC_{50}) in the range of 50-100 nM while an oligonucleotide targeted to the CAP region of the same message (I1751) has an IC_{50} in the range of 500 nM.

20 Figures 9 a and b are the nucleotide sequences of antisense oligonucleotides made in accordance with the teachings of the invention targeted to the transactivator and transrepressor regions of the E2 mRNA.

Figure 10 is a graphical depiction of the effects
25 of selected oligonucleotides targeted to the transactivator region of the E2 mRNA. The 15 to 20 mer antisense oligonucleotides made in accordance with the teachings of the invention are shown to inhibit E2 transactivation.

Figure 11 is a graphical depiction of the effects
30 of selected oligonucleotides targeted to the transrepressor region of the E2 mRNA. The 15 to 20 mer antisense oligonucleotides made in accordance with the teachings of the invention are shown to inhibit E2 transrepression.

Figure 12 is a photographic depiction of the consequences of reduction of E2 transactivator *in situ* on the biology of BPV-1. Antisense oligonucleotides made in accordance with the teachings of the invention were tested for

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the ability to inhibit or attenuate BPV-1 transformation of C127 cells. The photograph depicts petri dishes plated with test cells.

Figure 13 is a graphical depiction of the effects 5 of selected oligonucleotides targeted to the transactivator region of the E2 mRNA. The inhibition of BPV-1 focus formation by antisense oligonucleotides made in accordance with the teachings of the invention is depicted. These dose response curves for I1751 and I1753 show that I1753 had an 10 IC₅₀ in the range of 10 nM while I1751 had an IC₅₀ in the range of 100 nM.

Figure 14 is a graphical depiction of the effects 15 of antisense oligonucleotides made in accordance with the teachings of the present invention on the ability of BPV-1 to replicate its genome. Cells transformed by the virus were treated with I1753 and I1751 and the viral DNA quantitated.

Figure 15 shows the results of immunoprecipitation assays wherein metabolically labelled oligonucleotide-treated cells or untreated controls were immunoprecipitated using a 20 monoclonal antibody.

Figure 16 is the nucleotide sequence of HPV-11 in the region of the translation initiation codon of E2.

Figure 17 is a graph showing that ISIS 1751 and ISIS 25 1753 inhibited BPV-1 focus formation on C127 cells..

Figure 18 is a graph showing inhibition of HPV-11 E2-dependent transactivation in a concentration-dependent sequence-specific manner by ISIS 2105, having an IC₅₀ in the range of 5 μM.

SUMMARY OF THE INVENTION

30 In accordance with preferred embodiments of this invention, oligonucleotides and oligonucleotide analogs are provided which are hybridizable with messenger RNA from papillomavirus. This relationship is commonly denominated as "antisense." The oligonucleotides and oligonucleotide analogs 35 are able to inhibit the function of the RNA, either its translation into protein, its translocation into the

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cytoplasm, or any other activity necessary to its overall biological function. The failure of the messenger RNA to perform all or part of its function results in failure of the papillomavirus genome to be properly expressed; multiplication 5 fails, correct progeny are not formed in effective numbers, and the deleterious effects of those progeny upon animals infected with the papillomavirus are modulated.

It has now been found to be preferred to target portions of the papillomavirus genome, as represented by 10 certain of its mRNAs, for antisense attack. It has now been discovered that the E2, E1, E7 and E6-7 mRNAs of papillomaviruses are particularly suitable for this approach. Thus, it is preferred that the messenger RNA with which hybridization by the oligonucleotide is desired, be messenger 15 RNAs E2, E1, E7, or E6-7. In accordance with still more preferred embodiments, oligonucleotides are provided which comprise nucleotide base sequences designed to be complementary with RNA portions transcribed from the E2 transactivator region or the 5' common untranslated region of 20 the papillomavirus as exemplified, in bovine papillomavirus-1, by nucleotides 2443 through 3080 or 89 through 304.

In preferred embodiments, oligonucleotides are provided which are targeted to the mRNA CAP region and the translation codon for the E2 transactivator. These 25 oligonucleotides are designed to hybridize with E2 mRNA encoded by BPV-1 nucleotides 2443 through 4180.

A 20-mer phosphorothioate oligonucleotide, ISIS 2105 (SEQ ID NO: 7), targeted to the translation of initiation of both HPV-6 and HPV-11 E2 mRNA, covering nucleotides 2713 to 30 2732, is provided which inhibits E2-dependent transactivation in a concentration dependent sequence specific manner.

Methods of modulating the expression of papillomavirus have now been discovered comprising contacting messenger RNA from said papillomavirus with an oligonucleotide 35 hybridizable with a messenger RNA from the papillomavirus, which oligonucleotide inhibits the function of said messenger RNA when hybridized therewith. Employment of oligonucleotides

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or oligonucleotide analogs which are designed to hybridize with the E2, E1, E7, or E6-7 mRNAs of papillomavirus are preferred.

Inhibiting E2-dependent transactivation is most
5 preferred.

Additionally, methods of modulating the effects of a papillomavirus infection in an animal have now been discovered comprising contacting the animal with an oligonucleotide hybridizable with a messenger RNA from a
10 papillomavirus, that inhibits the function of said messenger RNA when hybridized therewith. Oligonucleotides hybridizable with E2, E1, E7, or E6-7 mRNAs of papillomavirus are preferred. Oligonucleotides targeted to the translation initiation of E2 mRNA is most preferred.

15 Diagnostics for detecting the presence or absence of papillomavirus employing such oligonucleotides or oligonucleotide analogs are also within this invention as are kits for such diagnostic activity and research reagents depending upon such hybridization.

20 **DETAILED DESCRIPTION OF THE INVENTION**

Genital warts are the most frequently diagnosed, viral, sexually transmitted disease. Clinically, they may be categorized into two major groups: condyloma acuminata and flat cervical warts. Condylomas have been shown to contain
25 virus particles and molecular studies have demonstrated that greater than 90% of these lesions contain either HPV-6 or HPV-11 DNA. Gissmann, L., Wolnik, L., Ikenberg, H., Koldovsky, U., Schnurch, H.G. & zur Hausen, H., Proc. Natl. Acad. Sci. USA 80:560-563 (1983). Condyloma acuminata
30 generally occur on the penis, vulva or in the perianal region. They may spontaneously regress or persist for years and progression to an invasive carcinoma occurs only at a low frequency. Unlike other genital warts, those occurring on the uterine cervix usually exhibit a flat rather than acuminate
35 morphology, and are usually clinically detected by Pap smear. A papillomavirus etiology for cervical dysplasia was suggested

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by the studies of cytologists in the late 1970s who demonstrated the association on Pap smear of cytologic changes due to HPV infection with those of dysplasia. Other studies showed the presence of viral particles and viral capsid antigen in some of the dysplastic cells of these lesions. This association was important because previous clinical studies had established that cervical dysplasia (also referred to as CIN, or cervical intra-epithelial neoplasia) was a precursor to carcinoma *in situ* which was in turn recognized to be a precursor to invasive squamous epithelial cell carcinoma of the cervix. HPV-types 16 and 18 were cloned out directly from cervical carcinoma. Dürst, M., Gissmann, L., Ikenberg, H. & zur Hausen, H., Proc. Natl. Acad. Sci. USA 80:3812-3815 (1983); Boshart, M., Gissmann, L., Ikenberg, H., Kleinheinz, A., Scheurlen, W. & zur Hausen, H., EMBO J. 3:1151-1157 (1984). These were subsequently used as hybridization probes to show that greater than 70% of the human cervical carcinomas and the derived cell lines scored positive for the presence of either of these HPV types. Another 20% contain additional HPV-types such as HPV-31, HPV-33, and HPV-35.

Data collected from the National Therapeutic Index showed that in 1984 there were 224,900 first office visits for genital warts and 156,720 first office visits for genital herpes. The incidence of genital warts has steadily increased throughout the 1970s and 1980s, as was recently demonstrated by an epidemiological study in which the mean incidence from 1950 to 1978 reached a peak of 106.5 per 100,000 population. The prevalence of cervical HPV infection in women aged 25 to 55 proved to be 0.8%, but in 22 year old women it was 2.7%. Recent studies on cytologically normal women have demonstrated the incidence of latent infection to be 11%. Thus, there appears to be a latent stage of the disease which suggests an even greater incidence and prevalence.

Active genital warts can be identified in approximately 2.5% of pregnant American women, thus being implicated in 60,000 to 90,000 pregnancies annually. HPV infections are more than twice as prevalent in pregnant women.

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Each year there are an estimated 1,500 new cases of laryngeal papillomatosis, indicating that the risk of infection from mother to newborn is 1:80 to 1:200.

Laryngeal papillomas are benign epithelial tumors of the larynx. Two PV types, HPV-6 and HPV-11, are most commonly associated with laryngeal papillomas. Clinically, laryngeal papillomas are divided into two groups, juvenile onset and adult onset. In juvenile onset it is thought that the neonate is infected at the time of passage through the birth canal of a mother with a genital PV infection. Disease is usually manifest by age 2 and is characterized by the slow but steady growth of benign papillomas that will ultimately occlude the airway without surgical intervention. These children will typically undergo multiple surgeries with the papillomas always reoccurring. Patients will ultimately succumb to complications of multiple surgery. To date there is no curative treatment for juvenile onset laryngeal papillomatosis and spontaneous regression is rare. Adult onset laryngeal papillomatosis is not as aggressive and will frequently undergo spontaneous remission.

The most common disease associated with papillomavirus infection are benign skin warts. Common warts generally contain HPV types 1, 2, 3, 4 or 10. These warts typically occur on the soles of feet, plantar warts, or on the hands. Common skin warts are most often found in children and young adults. Later in life the incidence of common warts decreases presumably due to immunologic and physiologic changes. Plantar warts can often be debilitating and require surgical removal and they frequently reoccur after surgery. To date there is no reliable treatment for plantar warts. Common warts of the hands are unsightly but rarely become debilitating and are therefore not usually surgically treated.

Epidermodysplasia verruciformis (EV) is a rare genetically transmitted disease which is characterized by disseminated flat warts that appear as small reddish macules. A variety of HPV types have been associated with EV. With time approximately one third of EV patients develop squamous

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cell carcinoma (SCC) of the skin at multiple sites. In general, SCC occurs on sun exposed areas of the skin. Only a subset of EV associated PV is consistently found in SCC, HPV-5 and HPV-8. Genetic predisposition, immunologic abnormalities, and UV irradiation as well as HPV may all contribute to the development of SCC in these patients.

The PV genome consists of a double stranded, covalently closed, circular DNA molecule of approximately 8,000 base pairs. The complete nucleotide sequence and 10 genetic organization of a number of animal and human PVs have been determined including bovine papillomavirus type 1 (BPV-1). Chen, E.Y., Howley, P.M., Levinson, A.D. & Seeburg, P.H., *Nature* 299:529-534 (1982). Schematic maps of several PV genomes are shown in Figure 1. Viral transcription is 15 unidirectional: all viral mRNAs are transcribed from the same strand of viral DNA. Engel, L.W., Heilman, C.A. & Howley, P.M., *J. Virol.* 47:516-528 (1983); Heilman, C.A., Engel, L., Lowy, D.R. & Howley, P.M., *Virology* 119:22-34 (1982). The coding strand contains 10 designated open reading frames 20 (ORFs). The individual ORFs have been classified as either "early" or "late" ORFs based on their position in the PV genome and their pattern of expression in non-productively versus productively infected cells. Figure 2 depicts the relationships of several ORFs for bovine papillomavirus-1.

25 Because of its ability to transform rodent cells and maintain its genome as an episome in transformed cells, BPV-1 has served as the model papillomavirus *in vitro* studies. As a result, BPV-1 is the best characterized of all the papillomaviruses. The BPV-1 genome is 7946 base pairs in 30 length and has been cloned and sequenced. Chen et al., 1982, *supra*. DNA sequence analysis of BPV-1 has defined 8 early (E) and 2 late (L) open reading frames (ORFs). Designation of ORFs as early or late was based on their pattern of expression in nonproductively infected transformed cells versus 35 permissively infected cells. Heilman et al., 1982, *supra*; Baker, C.C. & Howley, P.M., *EMBO J.* 6:1027-1035 (1987).

All ORFs are contained on the same strand of DNA and

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all mRNAs currently characterized have been shown to be transcribed from the coding strand. Amtmann, E. & Sauer, G., *J. Virol.* 43:59-66 (1982). The functions of the BPV-1 ORFs have been analyzed by recombinant DNA techniques and in vitro 5 cell culture systems. Several ORFs have been shown to have multiple functions. The E5 and E6 ORFs have been shown to encode transforming proteins. Yang, Y.C., Okayama, H. & Howley, P.M., *Proc. Natl. Acad. Sci. USA* 82:1030-1034 (1985). The E1 and E7 ORFs are involved in maintenance of high copy 10 number of the BPV-1 genome within the infected cell. Lusky, M. & Botchan, M.R., *J. Virol.* 53:955-965 (1985).

The 3' E1 ORF encodes a factor required for viral genome replication and maintenance of the viral genome. Lusky, M. & Botchan, M.R., *J. Virol.* 60:729-742 (1986). The 15 5' E1 ORF encodes a modular of viral DNA replication. Roberts, J.M. & Weintraub, H., *Cell* 46:741-752 (1986). The full length E2 ORF encodes a protein which transactivate viral transcription, (Spalholz, B.A., Yang, Y.C. & Howley, P.M., *Cell* 42:183-191 (1985)) while the 3' E2 ORF encodes a 20 transrepressor of viral transcription. Lambert, P.F., Spalholz, B.A. & Howley, P.M., *Cell* 50:69-78 (1987). No functions for E3, E4, and E8 of BPV-1 have yet been defined. L1 (Cowsert, L.M., Pilacinski, W.P. & Jenson, A.B., *Virology* 165:613-615 (1988)) and L2 encode capsid proteins.

25 In accordance with this invention, persons of ordinary skill in the art will understand that messenger RNA identified by the open reading frames of the DNA from which they are transcribed include not only the information from the ORFs of the DNA, but also associated ribonucleotides which 30 form regions known to such persons as the 5' cap region, the 5' untranslated region, and 3' untranslated region. Thus, oligonucleotide and oligonucleotide analogs may be formulated in accordance with this invention which are targeted wholly or in part to these associated ribonucleotides as well as to 35 the informational ribonucleotides.

Within the BPV-1 genome a region of about 1,000 base pairs in length, located between 7,100 and 8,100, has been

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identified that has no extensive coding potential. This region is referred to as the long control region (LCR). The LCR contains multiple CIS control elements that are critical for the regulation of viral transcription and viral 5 replication. A summary of functional assignments for the ORFs is set forth in Table 1.

Transcription of the BPV-1 genome is complicated by the presence of multiple promoters and complex and alternative splice patterns. Eighteen different mRNA species have been 10 identified so far by a variety of methods. Amtmann, E. & Sauer, G., *J.Virol.* 43:59-66 (1982); Burnett, S., Moreno-Lopez, J. & Pettersson, U., *Nucleic Acids Res.* 15:8607-8620 (1987); Engel, L.W., Heilman, C.A. & Howley, P.M., *J.Virol.* 47:516-528 (1983); Heilman, C.A., Engel, L., 15 Lowy, D.R. & Howley, P.M., *Virology* 119:22-34 (1982); Baker, C.C. & Howley, P.M., *EMBO J.* 6:1027-1035 (1987); Stenlund, A., Zabielski, J., Ahola, H., Moreno-Lopez, J. & Pettersson, U., *J. Mol. Biol.* 182:541-554 (1985); and Yang, Y.C., Okayama, H. & Howley, P.M., *Proc. Natl. Acad. Sci. USA* 82:1030-1034 20 (1985).

All early mRNAs appear to use a common polyadenylation signal at nucleotide (nt) 4180 which is positioned down stream of the early ORFs, while late mRNAs use a second polyadenylation signal at nt 7156. Sequence analysis 25 of BPV-1 cDNAs revealed the presence of multiple 5' splice sites (at nt 304, 864, 1234, 2505, 3764, and 7385) and 3' splice sites (at nt 528, 3225, 3605, 5609) resulting in alternative splicing events. The 5' end of most BPV-1 mRNAs map to nt 89 and contain a common region between nt 89 and the 30 first splice donor site at nt 304. The 5' end of other mRNAs map to nts 890, 2443 and 3080.

It is to be expected that differences in the DNA of papillomaviruses from different species and from different types within a species exist. It is presently believed, 35 however, that the similarities among the ORFs of the various PVs as the same might effect the embodiment of the present invention, outweigh the differences. Thus, it is believed,

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for example, that the E2 regions of the various PVs serve essentially the same function for the respective PVs and that interference with expression of the E2 genetic information will afford similar results in the various species. This is
5 believed to be so even though differences in the nucleotide sequences among the PV species doubtless exist.

Accordingly, nucleotide sequences set forth in the present specification will be understood to be representational for the particular species being described.
10 Homologous or analogous sequences for different species of papillomavirus are specifically contemplated as being within the scope of this invention.

Early genetic experiments showed that deletion or mutation of E2 resulted in loss of BPV focus forming activity
15 on C127 cells suggesting a transforming function for E2. Later studies showed that E2 was a regulator of viral transcription and that loss of transforming ability by mutation of E2 was due to the down regulation of other transforming genes through E2 conditional enhancers. The full
20 length E2 ORF encodes the E2 transactivator which stimulates transcription of viral early genes. The E2 transactivator is translated from an unspliced mRNA whose 5' end maps to nt 2443 as shown as species N in Figure 2. The E2 transrepressor is an N-terminally truncated form of the E2 transactivator,
25 generated by initiation of transcription within the E2 ORF by a promoter at nt 3089. Species O, Figure 2. The transactivating (5' portion) and DNA binding domains (3' portion) as well as the palindromic DNA recognition sequence (ACCN6GGT) of E2 have been identified. Both E2 mRNA and
30 protein have been shown to have a very short half life, on the order of less than about 60 minutes. The E2 transregulatory circuit is a general feature among papilloma viruses. E2 transregulator has been documented in every papillomavirus examined to date.

35 The PV genome is packaged in a naked icosahedral capsid 55 nm in diameter. The viral capsid is nonenveloped and is not glycosylated. Two viral encoded proteins,

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designated L1 and L2, make up the capsid. L1 is the major capsid protein, constitutes 80% of the protein present in the virion, and has a molecular weight ranging between 50 to 60 kD. The L2 is a minor capsid protein with a theoretical 5 molecular weight of 51 kD but has been shown to migrate at 76 kD. Because PV cannot be produced in vitro and very few mature virions are found in productive lesions, it has not been possible to do a detailed study of the serology of PV.

The papillomavirus life cycle is complex and poorly 10 understood at this time. To date, no in vitro system that allows production of mature PV virions has been developed, and as a result, it has not been possible to characterize the life cycle of papillomaviruses. PV have a restricted host range rarely crossing species barriers. In addition PV infect only 15 differentiating epithelium, either mucosal or keratinizing. Regulation of the PV gene expression is thought to be intimately linked to the differentiation program of host epithelial cells. The currently favored model of the PV life 20 cycle is as follows: infectious PV particles penetrate the outer layers of the epithelium via trauma and infect the basal cells of host tissue. There the virus is maintained as a relatively low copy extrachromosomal element. As the epithelial cells begin to undergo differentiation, the PV genome is replicated to high copy number, as the cells begin 25 to undergo the terminal stages of differentiation late genes are expressed and the viral genome is encapsulated.

Papillomavirus has been discovered to be an ideal target for antisense therapy. First, papillomavirus lesions are external, allowing topical approaches to delivery of 30 antisense oligonucleotides and eliminating many of the problems such as rapid clearance, and obtaining clinically active tissue concentrations of oligonucleotides associated with systemic administration of synthetic oligonucleotides. Second, the viral genome is maintained in the infected cell 35 as a separate genetic element. This opens the door to the possibility of curative therapy, as opposed to treatment of symptoms, by attacking replication functions of the virus.

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It has been discovered that the E2 ORF on papillomavirus genomes is particularly well-suited for antisense oligonucleotide design. E2 has been shown to be the major transactivator of viral transcription in both BPV-1 and 5 HPV systems. Mutations in the E2 ORF have pleiotropic effects on transformation and extrachromosomal DNA replication. DiMaio, D., *J. Virol.* 57:475-480 (1986); DiMaio, D. & Settleman, J., *EMBO J.* 7:1197-1204 (1988); Groff, D.E. & Lancaster, W.D., *Virology* 150:221-230 (1986); Rabson, M.S., 10 Yee, C., Yang, Y.C. & Howley, P.M., *J. Virol.* 60:626-634 (1986); and Sarver, N., Rabson, M.S., Yang, Y.C., Byrne, J.C. & Howley, P.M., *J. Virol.* 52:377-388 (1984). Subsequently, the E2 ORF has been shown to encode a transcriptional transactivator. Spalholz, B.A., Yang, Y.C. & Howley, P.M., 15 *Cell* 42:183-191 (1985). A truncated version of E2 created by initiation of translation at an internal AUG has been shown to be a transrepressor of transcription. Lambert, P.F., Spalholz, B.A. & Howley, P.M., *Cell* 50:69-78 (1987). Both the DNA binding domain, the carboxy terminal 100 amino acids, 20 (McBride, A.A., Schlegel, R. & Howley, P.M., *EMBO J.* 7:533-539 (1988)) and the DNA recognition sequence, (Androphy, E.J., Lowy, D.R. & Schiller, J.T., *Nature* 325, 70-73 (1987), and Moskaluk, C. & Bastia, D., *Proc. Natl. Acad. Sci. USA* 84:1215-1218 (1987)) have been identified. The E2 25 transcriptional regulatory circuit is a general feature among papillomaviruses. E2 transregulation has been documented in each of the other papillomaviruses examined to date. Gius, D., Grossman, S., Bedell, M.A. & Laimins, L.A., *J. Virol.* 62:665-672 (1988); Hirochika, H., Broker, T.R. & Chow, L.T., 30 *J. Virol.* 61:2599-2606 (1987); Chin, M.T., Hirochika, R., Hirochika, H., Broker, T.R. & Chow, L.T., *J. Virol.* 62:2994-3002 (1988); Phelps, W.C. & Howley, P.M., *J. Virol.* 61:1630-1638 (1987); and Thierry, F. & Yaniv, M., *EMBO J.* 6:3391-3397 (1987).
35 The inventors have determined that the identification of an obligatory viral transcription element that is shared among animal and human papillomaviruses causes

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E2 to be a prime target for an antisense approach towards papillomavirus research, diagnosis and therapy. In particular, a 20-mer phosphorothioate oligonucleotide, ISIS 2105 (SEQ ID NO: 7), designed to hybridize to the AUG region 5 of the HPV-11 E2 transactivator mRNA, covering nucleotides 2713 to 2732, was found to inhibit E2-dependent transactivation in a concentration dependent sequence specific manner.

The inventors have determined that the E1 locus is 10 also promising as a situs for attack upon papillomavirus. Initial mutational analysis of BPV-1 transformation of rodent fibroblast has identified the E1 as a candidate regulator of viral DNA replication. The 3' E1 ORF encodes a factor required for viral genome replication and maintenance. 15 Sarver, N., Rabson, M.S., Yang, Y.C., Byrne, J.C. & Howley, P.M., *J. Virol.* 52:377-388 (1984); Lusky, M. & Botchan, M.R., *J. Virol.* 53:955-965 (1985); and Lusky, M. & Botchan, M.R., *J. Virol.* 60:729-742 (1986). Inhibition of expression of E1 transcription is believed to be likely to inhibit the ability 20 of BPV-1 (and potentially HPV) to replicate their DNA in infected cells.

The inventors have also determined that the E7 site will likely provide a further entré to therapeutics, diagnostics and research into papillomavirus. In BPV-1, E7 25 has been shown to be involved in regulation of viral DNA replication. Berg, L.J., Singh, K. & Botchan, M., *Mol. Cell. Biol.* 6:859-869 (1986). In HPV-16, E7 has been demonstrated to be involved in transformation and immortalization. It is not clear at this time if the E7 of HPVs are involved in 30 replication of viral DNA, however, it is believed that E7 specific antisense oligonucleotides and analogs will inhibit replication of BPV-1 viral DNA.

The E6-E7 region of HPV has been found to be the transforming region. The exact role of this region in the 35 life cycle of the virus is unknown at this time. However, since this region plays a central in the biology of virally induced lesions antisense it has been determined that

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oligonucleotides targeted to this region are likely to be useful for the purposes of this invention as well.

Recent studies have suggest that antisense oligonucleotides directed towards the 5' regions of mRNAs and 5 preferably the cap region and the start codon are most effective in inhibiting gene expression. One feature of papillomavirus transcription is that many of the mRNAs have a common 5' untranslated region and cap. Thus it has been determined that antisense oligonucleotides directed towards 10 this region have the potential to incapacitate more than one mRNA. The shutting down of multiple viral genes will likely act at a minimum in an additive fashion and possibly synergistically in the eradication of the viral genome from the infected cell.

15 It will be appreciated that the ORFs of the papillomavirus genome which give rise to the mRNAs which are preferred targets for antisense attack in accordance with the practice of certain preferred embodiments of this invention also encode portions of other mRNAs as well.

20 The foregoing ORFs are summarized in Table 1.

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TABLE 1

Papillomavirus Open Reading Frames
and their assigned Functions

ORF	ASSIGNED FUNCTIONS
5 E (BPV-1)	(3'portion) Replication (BPV-1)
E2(full length)	Transcriptional transactivation (BPV-1, HPV-6, HPV-16)
3' portion)	Transcriptional repression (BPV-1)
E4	Cytoplasmic phosphoprotein in warts (HPV-1)
10 E5	Transformation, Stimulation of DNA synthesis (BPV-1)
E6	Transformation (BPV-1)
E7	Plasmid copy number control (BPV-1)
L1	Major capsid protein
15 L2	Minor capsid protein

The present invention employs oligonucleotides for use in antisense inhibition of the function of messenger RNAs of papillomavirus. In the context of this invention, the term "oligonucleotide" refers to a polynucleotide formed from naturally-occurring bases and cyclofuranosyl groups joined by native phosphodiester bonds. This term effectively refers to naturally-occurring species or synthetic species formed from naturally-occurring subunits or their close homologs. The term "oligonucleotide" may also refer to moieties which function similarly to oligonucleotides, but which have non naturally-occurring portions. Thus, oligonucleotides may have altered sugar moieties or inter-sugar linkages. Exemplary among these are the phosphorothioate and other sulfur containing species which are known for use in the art. In accordance with some preferred embodiments, at least one of the phosphodiester bonds of the oligonucleotide has been

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substituted with a structure which functions to enhance the ability of the compositions to penetrate into the region of cells where the RNA whose activity is to be modulated is located. It is preferred that such substitutions comprise
5 phosphorothioate bonds, methyl phosphonate bonds, or short chain alkyl or cycloalkyl structures. In accordance with other preferred embodiments, the phosphodiester bonds are substituted with structures which are, at once, substantially non-ionic and non-chiral, or with structures which are chiral
10 and enantiomerically specific. Persons of ordinary skill in the art will be able to select other linkages for use in the practice of the invention.

Oligonucleotides may also include species which include at least some modified base forms. Thus, purines and
15 pyrimidines other than those normally found in nature may be so employed. Similarly, modifications on the furanosyl portions of the nucleotide subunits may also be effected, as long as the essential tenets of this invention are adhered to. Examples of such modifications are 2'-O-alkyl- and 2'-halogen-
20 substituted nucleotides. Some specific examples of modifications at the 2' position of sugar moieties which are useful in the present invention are OH, SH, SCH₃, F, OCH₃, OCN, O(CH₂)_nNH₂ or O(CH₂)_nCH₃, where n is from 1 to about 10, and other substituents having similar properties.

25 Such oligonucleotides are best described as being functionally interchangeable with natural oligonucleotides or synthesized oligonucleotides along natural lines, but which have one or more differences from natural structure. All such analogs are comprehended by this invention so long as they
30 function effectively to hybridize with messenger RNA of papillomavirus to inhibit the function of that RNA.

The oligonucleotides in accordance with this invention preferably comprise from about 3 to about 50 subunits. It is more preferred that such oligonucleotides and analogs comprise
35 from about 8 to about 25 subunits and still more preferred to have from about 12 to about 20 subunits. As will be appreciated, a subunit is a base and sugar combination

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suitably bound to adjacent subunits through phosphodiester or other bonds. The oligonucleotides used in accordance with this invention may be conveniently and routinely made through the well-known technique of solid phase synthesis. Equipment 5 for such synthesis is sold by several vendors, including Applied Biosystems. Any other means for such synthesis may also be employed, however, the actual synthesis of the oligonucleotides is well within the talents of the routineer. It is also well known to prepare other oligonucleotide such 10 as phosphorothioates and alkylated derivatives.

The oligonucleotides of this invention are designed to be hybridizable with messenger RNA of papillomavirus. Such hybridization, when accomplished, interferes with the normal function of the messenger RNA to cause a loss of its utility 15 to the virus. The functions of messenger RNA to be interfered with include all vital functions such as translocation of the RNA to the situs for protein translation, actual translation of protein from the RNA, and possibly even independent catalytic activity which may be engaged in by the RNA. The 20 overall effect of such interference with the RNA function is to cause the papillomavirus to lose the benefit of the RNA and, overall, to experience interference with expression of the viral genome. Such interference is generally fatal.

In accordance with the present invention, it is 25 preferred to provide oligonucleotides designed to interfere with messenger RNAs determined to be of enhanced metabolic significance to the virus. As explained above, the E1, E2, E7, or E6-7 papillomavirus mRNAs are preferred targets. E2 is the most preferred target.

It is also preferred to interfere with RNA coded by 30 nucleotides substantially equivalent to nucleotides 2443 through 3080 or 89 through 304 of bovine papillomavirus-1 genome since these are believed to represent a particularly vulnerable situs for attack as they are believed to code for 35 a plurality of RNAs leading to essential proteins. It will be appreciated that differing papillomaviruses will have somewhat different structures from bovine papillomavirus-1,

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but that essentially similar areas may be routinely determined.

Figures 3, 4, and 5 represent these areas on the bovine papillomavirus-1 genome and any oligonucleotide or 5 oligonucleotide analog designed to interfere with RNA coded by their counterparts in particular papillomaviruses is likely to have especial utility in interfering with operation of those papillomaviruses. Exemplary oligonucleotides targeted at the E2 mRNA of bovine papillomavirus-1 are set forth in
10 Table 2.

TABLE 2
BPV-1 E2 antisense oligonucleotides

oligo-nucleotide	5'-----3'	Comments
001(LC001.AB1)	AGGTTTGCACCCGACTATGCAAGTACAAT	mRNA cap region
002(LC002.AB1)	TATGCCAAGTACAAT	mRNA cap region
003(LC003.AB1)	CCTTCGCGATGGCTCTCCATCCTCTCACT	initiation of translation
004(LC004.AB1)	GCATGCTGTCTCCAT	initiation of translation
005(LC005.AB1)	AAATGCGTCCAGCACCGGCCATGGTGCAGT	transrepressor start
006(LC006.AB1)	AGCACCGGCCATGGT	transrepressor start
007(LC007.AB1)	CAATGGCAGTGTGATCAGAACGTCAGCTGGC	translational termination
008(LC008.AB1)	GCAAGTGATCAGAAGT	translational termination
009(LC009.AB1)	ATTGCTGCAGCTTAACCATAATAATCTG	3' untranslated region
010(LC010.AB1)	CTTAAACCATAAAA	3' untranslated region
011(LC011.AB1)	AAAAAAAGATTTCACAATCTGCATCAGTAAT	5' untranslated region
012(LC012.AB1)	AAGATTTCACAATCTG	5' untranslated region
013(LC013.AB1)	CACTGTCCTAGGACAGTCACCCCTTTTTC	5' coding region
014(LC014.AB1)	GGACAGTCACCCCT	5' coding region
015(LC015.AB1)	TGTACAAATGGCTGTAGACAGTGTACCACT	mid coding region
016(LC016.AB1)	GCTGTAGACAGTGT	mid coding region
017(LC017.AB1)	GTGCGAGGGAGGACCGTCCCCGTACCCAACC	3' coding region
018(LC018.AB1)	GGACCGTCCGTACCC	3' coding region
019(LC019.AB1)	TTAACACAGGTGGAAATCCATCATGGTGGTG	5' coding region
020(LC020.AB1)	GGAATCCATCATGG	5' coding region

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Exemplary oligonucleotides targeted to the translation initiation codon of HPV-11 are set forth in Table 3.

TABLE 3

5 Antisense Oligonucleotides Targeted to the Translation
Initiation Codon of HPV-11

	<u>Compound</u>	<u>5'-----3'</u>	<u>SEQ ID NO:</u>
	I2100	GCTTCCATCTTCCTC	1
	I2101	GCTTCCATCTTCCTCG	2
10	I2102	TGCTTCCATCTTCCTCG	3
	I2103	TGCTTCATCTTCCTCGT	4
	I2104	TTGCTTCCATCTTCCTCGT	5
	I2105	TTGCTTCCATCTTCCTCGTC	6

Thus, it is preferred to employ any of the twenty
15 oligonucleotides set forth above or any of the similar
oligonucleotides which persons of ordinary skill in the art
can prepare from knowledge of the respective, preferred

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regions of the E2 ORF of a papillomavirus genome as discussed above. Similar tables may be generated for the other preferred ORF targets of papillomaviruses, E1, E7, and E6-7 from knowledge of the sequences of those
5 respective regions.

It is not necessary that the oligonucleotides be precisely as described in the foregoing table or precisely as required by a slavish interpretation of the mapping of the papillomavirus genome. Rather, the spirit of this
10 invention permits some digression from strict adherence to the genome structure and its literal "translation" into oligonucleotide. Modifications of such structures may be made, so long as the essential hybridizing function of the oligonucleotides results.

15 Similarly, it will be appreciated that species variation among the various papillomaviruses occur. While the various regions, e.g., E2, E1, etc., are very similar from species to species, some differentiation occurs. Alteration in the oligonucleotides to account for these
20 variations is specifically contemplated by this invention.

A preferred assay to test the ability of E2 specific antisense oligonucleotides to inhibit E2 expression was based on the well documented transactivation properties of E2. Spalholtz et al., J. Virol. 61:2128-2137 (1987). A
25 reporter plasmid (E2RECAT) was constructed to contain the E2 responsive element, which functions as an E2 dependent enhancer. E2RECAT also contains the SV40 early promoter, an early polyadenylation signal, and the chloramphenicol acetyl transferase gene (CAT). Within the context of this
30 plasmid, CAT expression is dependent upon expression of E2. The dependence of CAT expression on the presence of E2 has been tested by transfection of this plasmid into C127 cells transformed by BPV-1, uninfected C127 cells and C127 cells cotransfected with E2RECAT and an E2 expression vector.

35 Antisense oligonucleotides were designed to target the major E2 transactivator mRNA (Figure 6). Targets included, but were not limited to, the mRNA CAP region, the

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translation initiation codon, translation termination codon, and polyadenylation signal. Fifteen or 30 residue oligonucleotides complementary to the various targets were synthesized with a wild type phosphodiester

5 internucleosidic linkage or a modified phosphorothioate internucleosidic linkage. Oligonucleotides targeted to the mRNA CAP region (I1751) and the translation codon for the E2 transactivator (I1753) were shown to reduce E2 transactivation at 1 micromolar concentrations in

10 preliminary screens (Figure 7). Oligonucleotide I1756 targeted to the translation initiation codon of the E2 transrepressor (Figure 6) was able to give partial relief of transrepression as demonstrated by an increase in CAT activity (Figure 7). Other oligonucleotides of similar

15 length and base composition, but targeted to other areas of the E2 mRNA, as well as other nonsense control, failed to give an antisense effect. In general, oligonucleotides with the phosphorothioate internucleosidic linkage modification were more effective than oligonucleotides of

20 the same sequence containing the natural phosphodiester internucleosidic linkage. This is presumably due to the increased resistance of phosphorothioates to nucleases contained in the serum and within the cell. Dose response curves show that I1753 has an 50 % inhibitory concentration

25 (IC_{50}) in the range of 50 to 100 nM while I1751 has an IC_{50} ten fold higher in the range of 500 nM (Figure 8). After identification of the translation initiation codon of the E2 transactivator and transrepressor as successful antisense targets an additional set of phosphorothioates

30 were designed to more carefully probe the regions (Figure 9). These data showed that 15 to 20 mer oligonucleotides that covered the appropriate AUG could inhibit either E2 transactivation or transrepression (Figure 10 and 11).

In order to determine the consequences of reduction
35 of E2 transactivator *in situ* on the biology of the BPV-1, antisense oligonucleotides were tested for the ability to inhibit or attenuate BPV-1 transformation of C127 cells

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(Figure 12). Dose response curves for I1751 and I1753 showed that I1753 had an IC₅₀ in the range of 10 nM while I1751 had an IC₅₀ in the range of 100 nM (Figure 13) this 10 fold difference in the IC₅₀ of these two compounds in this 5 assay is similar to that observed in the inhibition of transactivation assay suggesting that the translation initiation codon is a better target.

To test the effect of E2 targeted antisense oligonucleotides on the ability of BPV-1 to replicate its genome, I-38 cells stably transformed by BPV-1 were treated with I1753 and I1751 and the viral DNA quantitated (Figure 14). After 48 hours of treatment at 1 micromolar concentration the viral DNA copy number on a per cell basis was reduced by factor of approximately 3. During the 10 course of this assay the cells divided between 2 and 3 times. This data suggests that the viral DNA failed to replicate synchronously with the cellular DNA.

In order to test the effect of I1753 on E2 protein synthesis, I-38 cells were metabolically labelled and 15 immunoprecipitated with an E2 specific monoclonal antibody. In cells not exposed to oligonucleotide or cells treated with sense or irrelevant oligonucleotides the 46 kd E2 protein is present (Figure 15). In cells treated with oligonucleotides targeted to the E2, the 46 kd band is lost 20 suggesting that the oligonucleotide is operating by hybridization arrest of translation.

The oligonucleotides and oligonucleotide analogs of this invention can be used in diagnostics, therapeutics and as research reagents and kits. For therapeutic use, the 25 oligonucleotide or oligonucleotide analog is administered to an animal suffering from a papillomavirus infection such as warts of the hands, warts of the feet, warts of the larynx, condylomata acuminata, epidermodysplasia verruciformis, flat cervical warts, cervical intraepithelial neoplasia, or any other infection involving a papillomavirus. It is generally preferred to apply the therapeutic agent in accordance with this invention

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topically or interlesionally. Other forms of administration, such as transdermally or intramuscularly, may also be useful. Inclusion in suppositories is presently believed to be likely to be highly useful. Use 5 of the oligonucleotides and oligonucleotide analogs of this invention in prophylaxis is also likely to be useful. Such may be accomplished, for example, by providing the medicament as a coating in condoms and the like. Use of pharmacologically acceptable carriers is also preferred for 10 some embodiments.

The present invention is also useful in diagnostics and in research. Since the oligonucleotides and oligonucleotide analogs of this invention hybridize to papillomavirus, sandwich and other assays can easily be 15 constructed to exploit this fact. Provision of means for detecting hybridization of oligonucleotide or analog with papillomavirus present in a sample suspected of containing it can routinely be accomplished. Such provision may include enzyme conjugation, radiolabelling or any other 20 suitable detection systems. Kits for detecting the presence or absence of papillomavirus may also be prepared.

EXAMPLES

EXAMPLE 1 Inhibition of Expression of BPV-1 E2 by Antisense Oligonucleotides

25 BPV-1 transformed C127 cells are plated in 12 well plates. Twenty four hours prior to transfection with E2RE1 cells are pretreated by addition of antisense oligonucleotides to the growth medium at final concentrations of 5, 15 and 30 mM. The next day cells are 30 transfected with 10 µg of E2RE1CAT by calcium phosphate precipitation. Ten micrograms of E2RE1CAT and 10 µg of carrier DNA (PUC 19) are mixed with 62 µl of 2 M CaCl₂ in a final volume of 250 µl of H₂O, followed by addition of 250 µl of 2X HBSP (1.5 mM Na₂PO₄, 10 mM KCl, 280 mM NaCl, 12 mM 35 glucose and 50 mM HEPES, pH 7.0) and incubated at room temperature for 30 minutes. One hundred microliters of

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this solution is added to each test well and allowed to incubate for 4 hours at 37°C. After incubation cells are glycerol shocked for 1 minute at room temperature with 15% glycerol in 0.75 mM Na₂PO₄, 5 mM KCl, 140 mM NaCl, 6 mM 5 glucose and 25 mM HEPES, pH 7.0. After shocking, cells are washed 2 times with serum free DMEM and refed with DMEM containing 10% fetal bovine serum and antisense oligonucleotide at the original concentration. Forty eight hours after transfection cells are harvested and assayed 10 for CAT activity.

For determination of CAT activity, cells are washed 2 times with phosphate buffered saline and collected by scraping. Cells are resuspended in 100 ul of 250 mM Tris-HCl, pH 8.0 and disrupted by freeze-thawing 3 times. 15 Twenty four microliters of cell extract is used for each assay. For each assay, the following are mixed together in an 1.5 ml Eppendorff tube: 25 µl of cell extract, 5 µl of 4 mM acetyl coenzyme A, 18 µl H₂O and 1 ul ¹⁴C-chloramphenicol, 40-60 mCi/mM and incubated at 37°C for 1 20 hour. After incubation chloramphenicol (acetylated and nonacetylated forms) are extracted with ethyl acetate and evaporated to dryness. Samples are resuspended in 25 µl of ethyl acetate and spotted onto a TLC plate and chromatograph in chloroform:methanol (19:1). TLC are 25 analyzed by autoradiography. Spots corresponding to acetylated and nonacetylated ¹⁴C-chloramphenicol are excised from the TLC plate and counted by liquid scintillation for quantitation of CAT activity. Antisense oligonucleotides that depress CAT activity in a dose dependent fashion are 30 considered positives.

**EXAMPLE 2 Inhibition of HPV E2 Expression
by Antisense Oligonucleotides**

The assay for inhibition of HPV E2 by antisense oligonucleotides is essentially the same as that for BPV-1 35 E2. For HPV assays appropriate HPVs are co-transfected into either CV-1 or A431 cells with PSV2NEO cells using the calcium phosphate method described above. Cells which take

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up DNA are selected for by culturing in media containing the antibiotic G418. G418 resistant cells are then analyzed for HPV DNA and RNA. Cells expressing E2 are used as target cells for antisense studies. For each antisense 5 oligonucleotide cells are pretreated as above followed by transfection with E2RE1CAT and analysis of CAT activity as above. Antisense oligonucleotides are considered to have a positive effect if they can depress CAT activity in a dose dependent fashion.

10 **EXAMPLE 3 Inhibition of HPV E7 Expression
by Antisense Oligonucleotides**

The E7 of HPV-16 has been shown to be capable of transactivating the Ad E2 promoter (Phelps, W. C. Yee, C. L., Munger, K., and Howley, P. M. 1988, The Human 15 Papillomavirus Type 16 E7 Gene Encodes Transactivation and Transformation Functions Similar to Those of Adenovirus E1A, Cell 53:539-547. To monitor this activity, a plasmid is constructed which contained the chloramphenicol transferase gene under the control of the Ad E2 promoter 20 (AdE2CAT). Under the conditions of this assay, CAT expression is dependent on expression of HPV E7. For this assay, cell lines are developed that contain the HPV E7 under the control of the SV40 early promoter. For each antisense oligonucleotide, cells are pretreated as above 25 followed by transfection with AdE2CAT and analysis of CAT activity as above.

**EXAMPLE 4 Inhibition of Expression of BPV-1
E1 by Antisense Oligonucleotides**

The E1 of BPV-1 has been shown to be a regulator of 30 viral genome replication. To test the effects of antisense oligonucleotides on viral replication C127 cells infected with BPV-1 are treated with E1 specific antisense oligonucleotides by addition of oligonucleotides to the growth medium at final concentrations of 5, 15 and 30 μ M. 35 The effects of the oligonucleotides are evaluated by a routine Northern blot analysis for quantitation of both E1 specific RNA as well as total viral RNA. In addition, the effects of antisense oligonucleotides on viral genome copy

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number are determined by Southern blot on total genomic DNA.

**EXAMPLE 5 Determination of Efficacy of BPV-1
Antisense Oligonucleotides on
Experimentally Induced Bovine Fibropapillomas**

5 Multiple bovine fibropapillomas are induce on calves by direct infection of the epidermis with purified BPV-1. Upon development, fibropapillomas are treated with oligonucleotides that had positive results in vitro as well 10 as controls. Antisense oligonucleotides that induce regression of the fibropapilloma are considered as positives.

**EXAMPLE 6 Design and Synthesis of
Oligonucleotides Complementary to E2 mRNA**

15 Antisense oligonucleotides were designed to be complementary to various regions of the E2 mRNA as defined by the published nucleotide sequence of BPV-1 (Chen, E. Y., Howley, P. M., Levinson, A. D., and Seenburg, P. H., The primary structure and genetic organization of the bovine 20 papillomavirus type 1 genome, *Nature* 299:529-534 (1982)) and cDNA structure of the major E2 transactivator mRNA (Yang, Y. C., Okayama, H., and Howley, P. M., Bovine papillomavirus contains multiple transforming genes, *Proc. Natl. Acad. Sci. USA* 82:1030-1034 (1985)). Antisense 25 oligonucleotides targeted to the translation initiation codon of HPV-11 E2 were based on the published sequence of HPV-11 (Dartmann, K., Schwarz, E., Gissamnn, L., and zur Hausen, *Virology* 151:124-130 (1986)). Solid-phase oligodeoxyribonucleotide syntheses were performed using an 30 Applied Biosystems 380B automated DNA synthesizer. For the phosphorothioate oligonucleotides, sulfurization was performed after each coupling using 0.2 M ^{35}S -1,2-Benzodithiol-3-one-1,1-dioxide dissolved in acetonitrile as described by Iyer et al. (Iyer, R. P., Phillips, L. R., 35 Egan, W., Regan, J. and Beauchage, S. L., The Automated Synthesis of Sulfur-Containing Oligodeoxyribonucleotides Using ^{35}S -1,2-Benzodithiol-3-one 1,1-Dioxide as a Sulfur-Transfer Reagent, *J. Org. Chem.* 55:4693-4699 (1990)). To

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insure complete thioation, the growing oligonucleotide was capped after each sulfurization step. After cleavage from the synthesis matrix, deprotection and detriylation oligonucleotides were ethanol precipitated twice out NaCl
5 and suspended in water. The concentration of oligonucleotide was determined by optical density at 260 nm. For use in cell culture assays, oligonucleotides were routinely diluted to 100 micromolar stocks and stored at - 80°C until use. The purity, integrity, and quantity of the
10 oligonucleotide preparations were determined by electrophoresis on 20% acrylamide 7 M urea gels (40 cm x 20 cm x 0.75 mm) prepared as described by Maniatis et al.
(Maniatis, T., Fritsch, E. F. and Sambrook, J. Molecular Cloning: A Laboratory Manual, Cold Spring Harbor
15 Laboratory, New York, 1982). Electrophoresed oligonucleotides were visualized within the gel by staining with "Stains-all" , 1-ethyl-2[3-(1-ethylnaphthol[1,2-d]-thiazolin-2-ylidene)-2-Methyl-Propenyl[naphthol[1,2d]-thiazolium bromide purchased from Sigma, E-9379, (Dahlberg,
20 A. E., Digman, C. W. and Peacock, A. C., *J. Mol. Biol.* 41:39 (1969)).

EXAMPLE 7 Molecular Constructs

The E2 chloramphenicol acetyl transferase (CAT) reporter plasmid used in this study has been previously
25 described (Spalholz, B. A., Byrne, J. C. and Howley, P. M., Evidence for Cooperativity between E2 Binding Sites in E2 trans-regulation of Bovine Papillomavirus Type 1, *J. Virol.* 62:3143-3150 (1988)). Briefly, the E2 responsive element, E2RE1, (nt 7611-7806) of BPV-1 was reconstructed using
30 oligonucleotides and cloned into pSV2CAT that had been deleted of the SV40 enhancer, SphI fragment. Expression of CAT from this plasmid has been shown to be dependent upon full length E2. Plasmid C59 contain an E2 cDNA expressed from the simian virus 40 promoter and enhancer and has been
35 described in detail elsewhere (Yang, Y.-C., Okayama, H. and Howley, P. M., Bovine papillomavirus contains multiple transforming genes, *Proc. Natl. Acad. Sci. USA* 82:1030-1034

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(1985)). Two HPV-11 full length E2 expression constructs were made. IPV115 contains the XmnI fragment of HPV-11 (nt 2665-4988) cloned into the SmaI site of pMSG purchased from Pharmacia (catalog number 27-4506), IPV118 contains the 5 same HPV-11 XmnI fragment cloned into the SmaI site of pSVL (Pharmacia, catalog number 27-4509).

EXAMPLE 8 Cell Lines

Mouse C127 cells (Dvoretzky, I. Schober, R., and Lowy, D., Focus Assay in Mouse Cells for Bovine Papillomavirus type 1, *Virology* 103:369-375 (1980)) were grown in Dulbecco's Modified Eagle's medium supplemented with 10% fetal bovine serum, penicillin (100U/ml), streptomycin (100ug/ml), and L-glutamine (4 mM). I-38 cell line was derived from a single focus of C127 cells 15 transformed by purified BPV-1 (Cowser, L. M., Lake, P., and Jenson, A. B., Topographical and conformational Epitopes of Bovine Papillomavirus type 1 Defined by Monoclonal Antibodies, *JNCI* 79:1053-1057 (1987)).

EXAMPLE 9 Oligonucleotide Inhibition of E2 Dependent Transactivation Assays

To test an oligonucleotide's ability to inhibit E2 transactivation or transrepression, I-38 cells were plated at 1×10^4 cells per cm^2 in 60 mm petri dishes 24 hours before transfection. Sixteen hours prior to transfection, 25 media was aspirated and replaced with media containing oligonucleotide at the appropriate concentration. One hour prior to transfection, media was aspirated and replaced with fresh media without oligonucleotide. Cells were transfected by the calcium phosphate precipitation method 30 as described by Graham et al. 1973 (Graham, F. L. and van der Eb, A. J., A New Technique for the Assay of Infectivity of Human Adenovirus 5 DNA, *Virology* 52:456-461 (1973)) with a total of 20 micrograms of DNA in one milliliter of precipitate. Each 60 mm dish received 200 microliters of 35 precipitate containing 4 micrograms of DNA. Four hours after the addition of precipitated DNA, the supernatant was aspirated and the cells treated with 15% glycerol (Frost, E. and Williams, J., Mapping Temperature-Sensitive and

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host-range mutation of Adenovirus type 5 by Marker Rescue, *Virology* 91:39-50 (1978)). After washing, cells were refed with media containing oligonucleotide at the original concentration and incubated for 48 hours.

5 **Example 10 Antisense Oligonucleotide Inhibition of Focus Formation**

The ability of antisense oligonucleotides that inhibited E2 transactivation to inhibit viral focus formation, a measure of transformation, was tested. Mouse 10 C127 cells were plated at subconfluence (5×10^4 cells/cm²) in 60 mm petri dishes. Cells were either infected with 50 focus forming units (FFU) per plate of purified BPV-1 or transfected with cloned BPV-1 DNA. Twenty-four hours after infection or transfection, oligonucleotides were added to 15 the medium. Medium was changed every 72 hours with fresh oligonucleotide added with each change. Twenty-five days post infection, cells were fixed in 10% formalin in PBS for 5 minutes and stained with 0.14% methylene blue aqueous solution for 10 minutes. Plates were washed with water and 20 foci counted.

The two compounds that inhibited E2 transactivation, ISIS 1751 and ISIS 1753, also inhibited BPV-1 focus formation on C127 cells (Figure 17).

This effect was found to be concentration-dependent 25 with an IC₅₀ of 10 nM for ISIS 1753. A clear IC₅₀ for ISIS 1751 was not achieved, but appears to be in the range of 500 to 1000 nM. ISIS 1755, which increased E2-dependent transactivation, had no effect on BPV-1 focus formation.

30 **Example 11 Antisense Oligonucleotide Inhibition of Human Papillomavirus HPV-11 E2 Transactivation**

Based on the BPV results obtained, a twenty nucleotide phosphorothioate oligonucleotide, ISIS 2105, was designed to hybridize to the AUG (translation initiation) region of the HPV-11 E2 transactivator mRNA. For 35 inhibition of HPV-11 E2 transactivation, C127 cells were pretreated with oligonucleotide by addition to the medium. The next day, medium was aspirated and replaced with fresh medium without oligonucleotide. Cells were co-transfected

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with 2 µg IPV 118 HPV-11 E2 expression plasmid, 2 µg IPV120-15 D2-CAT reporter plasmid, and 2 µg PCH110. Following transfection, cells were treated again with oligonucleotide and incubated for 48 hours. Cells were 5 harvested and processed for CAT and β-galactosidase assays. Chloramphenicol acetyltransferase activity was determined using standard protocols [Groman, C.M., Moffat, L.F., and Howard, B.H., *Mol. Cell. Biol.* 2:1044-1051 (1982)]. Acetylated and nonacetylated reaction products were 10 separated by thin layer chromatography and quantitated using a Molecular Dynamics PhosphoImager (Molecular Dynamics, Sunnyvale CA). β-galactosidase activity was determined using standard methods [Herbomel et al., *Cell* 39:653 (1984)].

15 ISIS 2105 was found to inhibit HPV-11 E2-dependent transactivation in a concentration-dependent sequence-specific manner with an IC₅₀ in the range of 5 µM (Figure 18). ISIS 2324, a BPV-1 E2-specific oligonucleotide, did not inhibit HPV-11 E2 transactivation significantly. This 20 is expected based on lack of sequence homology.

**Example 12 Inhibition of Human Cancer Cells
by HPV-18 Antisense Oligonucleotides**

HPV is implicated in both oral cancer and cervical cancer in humans. Synthetic oligonucleotides were made 25 that corresponded to the start codon regions of the E6 and E7 genes of HPV-18. The oligonucleotides are shown below:

<u>Compound</u>	<u>Sequence</u>	<u>Gene</u>	<u>Target region</u>	<u>SEQ ID NO:</u>
ISIS 2490	AGCGCGCCATAGTATTGTGG	E6	Initiation of translation	8
30 ISIS 2491	GTCCATGCATACTTAATATT	E7	Initiation of translation	9
ISIS 2494	TATTACGTACTAGATTCTAC		Nonsense control	10

The HPV-18-transformed oral cancer cell line 1483 and the cervical cancer cell line C4-1 were used, both of 35 which contain HPV-18 DNA. Cells were plated on day 1;

- 34 -

after cells attached, medium was aspirated and replaced with fresh medium containing 2 μ M or 5 μ M oligonucleotide. Medium was aspirated and replaced with medium containing fresh oligonucleotide on day 3. Replicate plates were 5 harvested on days 2, 3, 4, 5 and 6, and the cells were counted. Data are plotted as total cells per plate and are the average of triplicates.

By the tenth day of culture, 2 μ M ISIS 2491 had reduced the growth of the 1483 cells to 45% of cells 10 treated with random sequence oligonucleotide control, while 5 μ M ISIS 2491 reduced it to 18%. A combination of 2 μ M ISIS 2490 and 2 μ M 2491 reduced growth of the 1483 cells to 3% of the control. The combination of oligonucleotides reduced the C4-1 cells by 89% by the sixth day of culture. 15 Morphological examination showed that neither ISIS 2490 or 2491 alone produced significant changes in the 1483 cells apart from reduced growth, while the combination of oligonucleotides caused rounding and loss of attachment. These results support a role for antisense molecules in 20 regulation of growth of cancer cells that carry human papillomavirus.

While a number of specific embodiments have been set forth, the present invention is to be limited only in accordance with the following claims.

- 35 -

SEQUENCE LISTING

(1) GENERAL INFORMATION:

(i) APPLICANT: Crooke, Stanley T., Mirabelli,
Christopher K., Ecker, David J., Cowser, Lex M.

(ii) TITLE OF INVENTION: ANTISENSE OLIGONUCLEOTIDE
INHIBITORS OF PAPILLOMAVIRUS

(iii) NUMBER OF SEQUENCES: 10

(iv) CORRESPONDENCE ADDRESS:

(A) ADDRESSEE: WOODCOCK WASHBURN KURTZ

MACKIEWICZ & NORRIS

(B) STREET: One Liberty Place - 46th Floor

(C) CITY: Philadelphia

(D) STATE: Pennsylvania

(E) COUNTRY: USA

(F) ZIP: 19103

(v) COMPUTER READABLE FORM:

(A) MEDIUM TYPE: DISKETTE, 3.5 INCH, 1.44 Mb

STORAGE

(B) COMPUTER: IBM PS/2

(C) OPERATING SYSTEM: PC-DOS

(D) SOFTWARE: WORDPERFECT 5.0

(vii) PRIOR APPLICATION DATA:

(A) APPLICATION NUMBER: n/a

(B) FILING DATE: herewith

(viii) ATTORNEY/AGENT INFORMATION:

(A) NAME: Jane Massey Licata, Esquire

(B) REGISTRATION NUMBER: 32,257

(C) REFERENCE/DOCKET NUMBER: ISIS-0285

(ix) TELECOMMUNICATION INFORMATION:

- 36 -

(A) TELEPHONE: (215) 568-3100

(B) TELEFAX: (215) 568-3439

(2) INFORMATION FOR SEQ ID NO: 1:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 15

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(iv) ANTI-SENSE: yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

GCTTCCATCT TCCTC 15

(2) INFORMATION FOR SEQ ID NO: 2:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 16

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(iv) ANTI-SENSE: yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2:

GCTTCCATCT TCCTCG 16

(2) INFORMATION FOR SEQ ID NO: 3:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 17

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(iv) ANTI-SENSE: yes

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

TGCTTCCATC TTCCTCG 17

(2) INFORMATION FOR SEQ ID NO: 4:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 18
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(iv) ANTI-SENSE: yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 4:

TGCTTCCATC TTCCTCGT 18

(2) INFORMATION FOR SEQ ID NO: 5:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 19
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(iv) ANTI-SENSE: yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 5:

TTGCTTCCAT CTTCCCTCGT 19

(2) INFORMATION FOR SEQ ID NO: 6:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(iv) ANTI-SENSE: yes

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 6:

TTGCTTCCAT CTTCCTCGTC 20

(2) INFORMATION FOR SEQ ID NO: 7:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(iv) ANTI-SENSE: yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:

TTGCTTCCAT CTTCCTCGTC 20

(2) INFORMATION FOR SEQ ID NO: 8:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(iv) ANTI-SENSE: yes

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:

AGCGCGCCAT AGTATTGTGG 20

(2) INFORMATION FOR SEQ ID NO: 9:

(i) SEQUENCE CHARACTERISTICS:

- (A) LENGTH: 20
- (B) TYPE: nucleic acid
- (C) STRANDEDNESS: single
- (D) TOPOLOGY: linear

(iv) ANTI-SENSE: yes

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:

GTCCATGCAT ACTTAATATT 20

(2) INFORMATION FOR SEQ ID NO: 10:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 20

(B) TYPE: nucleic acid

(C) STRANDEDNESS: single

(D) TOPOLOGY: linear

(iv) ANTI-SENSE: no

(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:

TATTACGTAC TAGATTCTAC 20

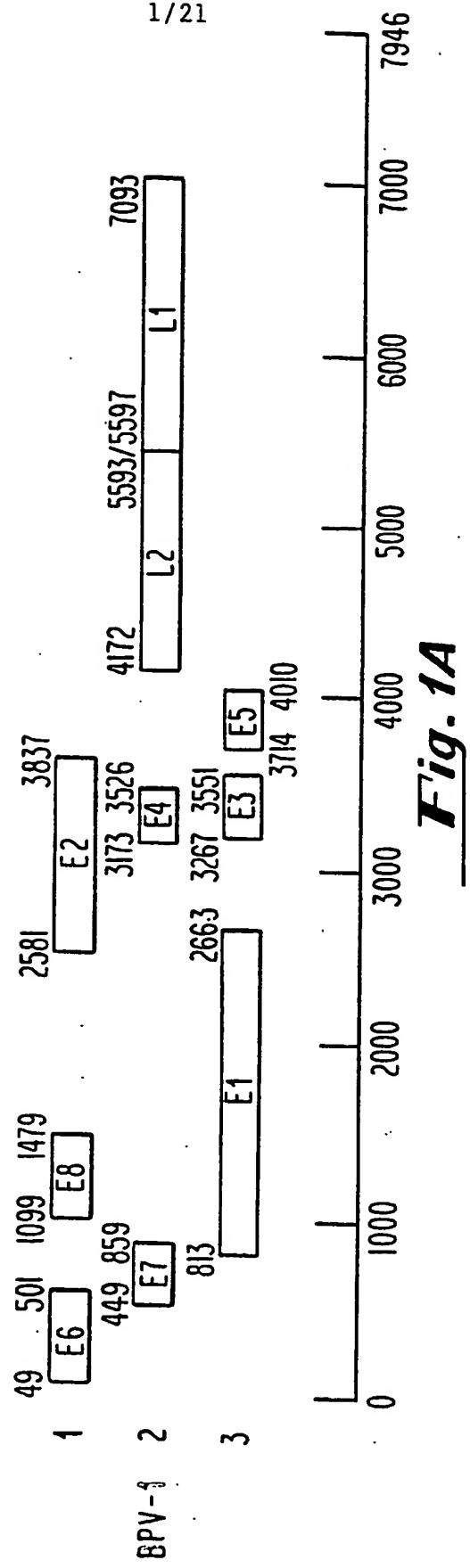
- 40 -

CLAIMS

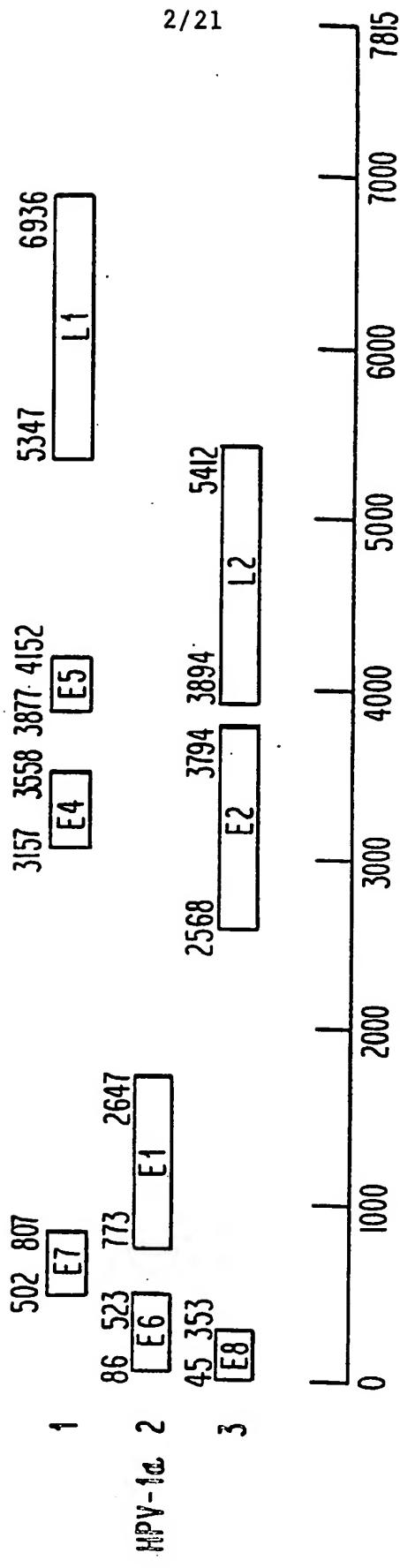
What is claimed is:

1. An antisense oligonucleotide comprising from 8 to 50 nucleotides, hybridizable with the translation initiation region of a human papillomavirus E2 transactivator messenger RNA that inhibits the function of said messenger RNA when hybridized therewith.
2. An antisense oligonucleotide of claim 1 having at least one phosphorothioate internucleotide bond.
- 10 3. An antisense oligonucleotide having SEQ ID NO: 7.
4. An antisense oligonucleotide comprising from 8 to 50 nucleotides, hybridizable with the translation initiation region of the E6 or E7 messenger RNA of a human papillomavirus that inhibits the function of said messenger RNA when hybridized therewith.
- 15 5. An antisense oligonucleotide of claim 4 having at least one phosphorothioate internucleotide bond.
6. An antisense oligonucleotide of claim 5 having SEQ ID NO: 8 or SEQ ID NO: 9.
- 20 7. A method of regulation of growth of cancer cells that carry human papillomavirus comprising contacting cancer cells that carry human papillomavirus with an antisense oligonucleotide of claim 4.
- 25 8. A method of regulation of growth of cancer cells that carry human papillomavirus comprising contacting cancer cells that carry human papillomavirus with an antisense oligonucleotide of claim 5.
9. A method of regulation of growth of cancer 30 cells that carry human papillomavirus comprising contacting cancer cells that carry human papillomavirus with an antisense oligonucleotide of claim 6.

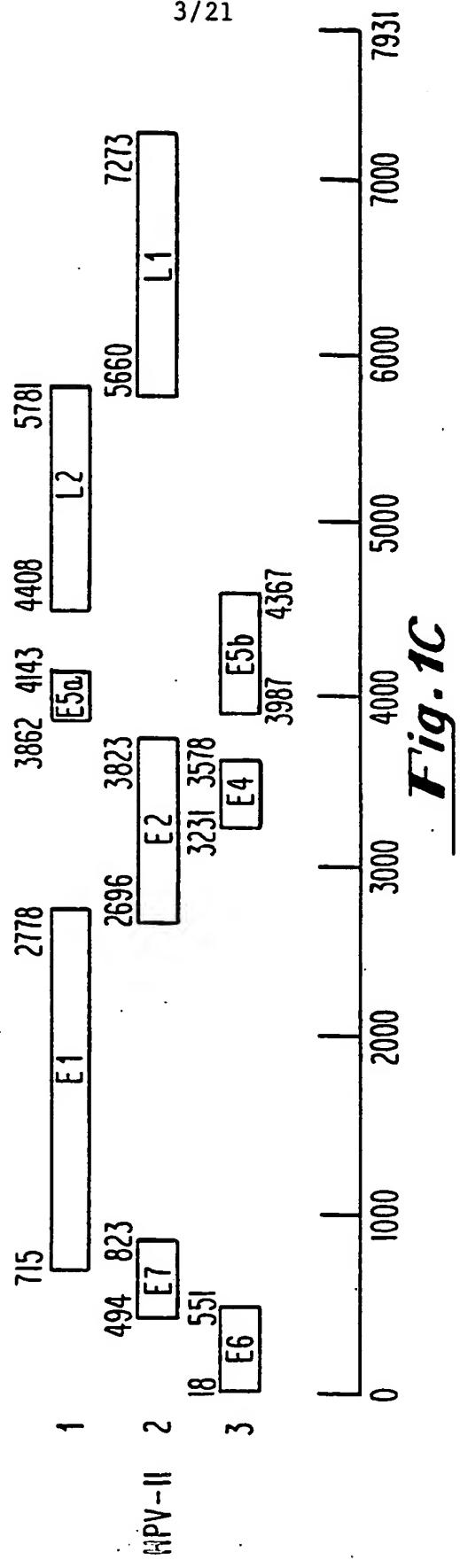
1/21



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***Fig. 1B***

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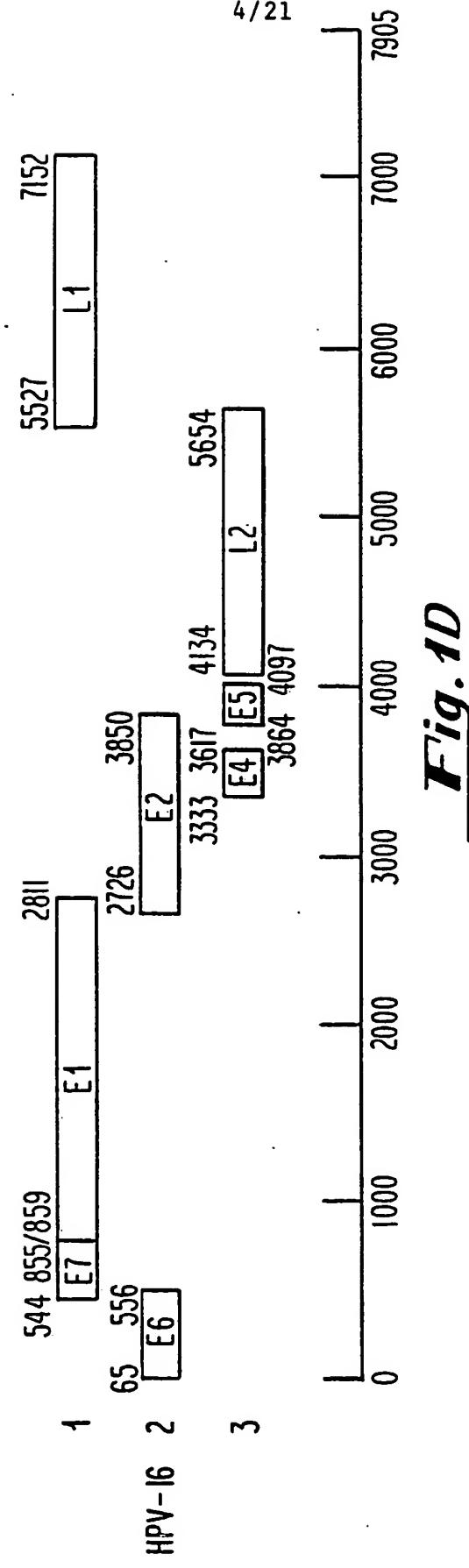
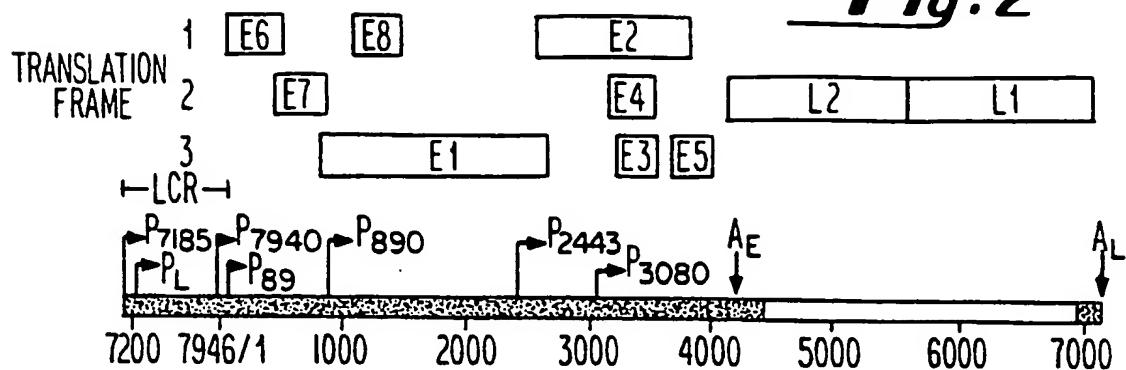


Fig. 1D

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Fig. 2

<u>SPECIES</u>	<u>FIRST POTENTIAL CODING ORF</u>		
A 7185	4203	?	
B 7879	864	3225	E6
C 89	304	3225	E6/E4
D 89	(304)	4203	?
E 89	304 528 864	3225	E6/E7
F 89	(864)	(3225)	E6
G 89	(304)(528)	4203	(E6/E7)
H 89	4203	E6	
I 138	304 528 1235	3225	E6/E7
J 890	1235	3225	E8/E2
K	(1235)	(3225)	?
L	(2505) (3225)	4203	?
M	2443 2505 3225	4203	E5
N	2443	4203	E2
O	3080	4203	E2
P 7255	4203	E6	
Q 7232	864	3225	E6
R 7246	7385	3225	E4
S 7217	7385	3605	E2
T 7269	7385	3605 3764	L1
U	4096	5609 L2	7175

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Fig. 3A

1 ATTGTACTT GCATAGTCGG GTGCCAACCT TTCGCTTGTCA GCAGCCATGC ACAGATGAAT
 61 CGGGTGAGCA ACCTTTTAAT ATTACTGATG CAGATTGAA ATCTTTTTT GTAAGGTAT
 121 GGGGGTTT AGACCTGATT GACGAGGAGG AGGATACTGAG AGAGGATGGAG CACAGCATGC
 181 GAACTGTTAC ATGTAGCGCA AGAAACACAA ATGCAGTGTAG TGAGAAAAG TAGTGATAAG
 241 TTGCAAGATC ATATACTGTA CTGGACTGCT GTTAGAACTG AGAACACACT GCTTTATGCT
 301 GCAAGGAAAA AAGGGGTGAC TGTCCCTAGGA CACTGCGAGG TACACACCT TGTAGTTGT
 361 CAAGAGAG CCAAGCAGGC CATTGAATG CAGTTGTCTT TGCAAGGAGT AGCAAAACT
 421 GAGTTTGGG ATGAACCAGT GTCTTTGCTT GACACAAGCT GGACACGATA TATGTCAAGA
 481 CCTAAACGGT GCTTTAAGAA AGGCGCCAGG GTGGTAGAGG TGAGTTGAA TCGAAATGCA
 541 AGCAATACAA ACTGGTACAC TGTCTACAGC AATTTGTACA TGGGCACAGA GGACGGCTGG
 601 CAGCTTGCGA AGGCTGGGC TGACCGGAACG GGGCTCTACT ACTGCACCAT GCCCGGTGCT
 661 GGACGCCATT ACTATTCTCG CTTTGGTGTAC GAGGCAAGCCA GATTTAGTAC AACAGGGCAT
 721 TACTCTGTAAGGATCAGGAGG CAGAGTGTAT GCTGGTGTCT CATCCACCTC TTCTGATTT
 781 AGAGATGCC CAGACGGAGT CTGGGTGCA TCCGAAGGAC CTGAAGGAGA CCCTGCAAGGA
 841 AAAGAAGCCG AGCCAGCCC GCCTGTCTCT TCCTTGCTCG GCTCCCCCGC CTGGGGTCCC
 901 ATCAGAGCCG GCCTCGGTG GGTAACGGAC GGTACGGCT GGTCCTCGCT CGCACCCCTA CAATTTCCT
 961 GCAGGCTCGG GGGGCTCTAT TCTTCGGCTCT TCCTCCACCC CGTGCAGGG CACGGTACCG
 1021 GTGGACTTGG CATCAAGGCC GGAAGAAGAG GAGCAAGTGC CGAAGTCCAC AGAGGAAGAA
 1081 CCAGTGACTC TCCCAAGGG CACCAAAAT GATGGATTCC ACTGTAAAGGACGGGG
 1141 TCATGCTTGT CTCTAATTTC AGGAACACTGCT ACCAGGTAAGTGCATG CTTTCGGGTG
 1201 AAAAGAACC ATAGACATCG CTACGAGAAC TGCAACCCA CCTGGTTCAC AGTTGCTGAC
 1261 AACGGGTGCTG AAAGACAAGG ACAAGCACAA ATACTGATCA CCTTTGGATC GCCAAGTCAA

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1321 AGGCAAGACT TTCTGAAACA TGTACCACTA CCTCCCTGGAA TGAACATTCA CGGCTTTACA
1381 GCCAGCTTGG ACTTCTGATC ACTGCCATTG CTTTTCTTC ATCTCACCTGG TGTACTATGC
1441 CAAATCTATG GTTCTTATTG TTCTGGAC TAGTTGCTGC ATGCCAACTG CTGCTATTAC
1501 TGTTCTTACT CTTGTTTTT CTTGTATACT GGGATCATTT TGAGTGCCTCC TGTACAGGTC
1561 TGCCTTTA ATGCCTTAC ATCACTGGCT ATTGGCTGTG TTCATTGTG TTTTACTGT TGTGTGGATT
1621 TGATTGTT TATATACTGT ATGAAGTTT TGTTTGTGCT CTTGTATTGC TGTTGTAAAG
1681 TTTTTACTA GAGTTTGAT TCCCCCTGCT CAGATTAT ATGGTTTAAG CTGCAGCAAT
1741 AAAATGAGT GCACGAAAAA G

Fig. 3B

1 ATTGTACTT GCATAGTCGG GTGCAAACCT TTCGCTTGA GCAGCCATGC ACAGATGAAT
 61 CGGGTACCA ACCTTTAAT ATTACTGATC CAGATTGAA ATCTTTTTT GTAAGGTTAT
 121 GGGGGCTT AGACCTGATC GACGAGGG AGGATAGTGA AGAGGATGGA GACAGCATGC
 181 GAACGTTAC ATGTAGCGCA AGAACACAA ATGCAGTTGA TTGAGAAAAG TAGTGATAAG
 241 TTGCAAGATC ATATACTGTA CTGGACTGCT GTTAACTG AGAACACACT GCTTTATGCT
 301 GCAAGGAAA AAGGGGTGAC TGTCCCTAGGA CACTGCAGAG TACCACACTC TGTAAGTTGT
 361 CAAGAGAG CCAAGCAGGC CATIGAAATG CAGTTGTCT TGCAAGGTT AAGCAAAACT
 421 GAGTTGGGG ATGAAACCATG GTCTTTGCTT GACACAAGCT GGGACCGATA TATGTCAGAA
 481 CCTAAACGGT GCTTAAAGAA AGGCCAGG GTGGTAGAGG TGGAGTTGA TGGAAATGCA
 541 AGCAATACAA ACTGGTACAC TGTCTACAGC AATTGTACA TGGCACAGA GGACGGCTGG
 601 CAGCTTGCAGA AGGCTGGGGC TGACGGAAC TGGCTCTA

Fig. 4

1 ACATGGACCT GAAACCTTT GCAAGAACCA ATCCATTCTC AGGGTTGGAT TGTCTGTGGT
 61 GCAGAGGCC TCTTACAGAA GTTGATGCTT TTAGGTGCAT GGTCAAGAC TTTCATGTTG
 121 TAATTGGGA AGGCTGTAGA TATGTTGCAT GTACCCATTG TCTGAAAC TGTTTAGCTA
 181 CTGAAAGAAC ACTFTGGCAA GGTGTTCCAG TAACAG

Fig. 5

Oligo	5'	3'	Comments
001	011	LC001.ABI	mRNA cap region
		LC002.ABI	mRNA cap region
		LC003.ABI	initiation of translation
		LC004.ABI	initiation of translation
		LC005.ABI	transrepressor start
		LC006.ABI	transrepressor start
		LC007.ABI	translational termination
		LC008.ABI	translational termination
		LC009.ABI	3' untranslated region
		LC010.ABI	3' untranslated region
		LC011.ABI	5' untranslated region
		LC012.ABI	5' untranslated region
		LC013.ABI	5' coding region
		LC014.ABI	5' coding region
		LC015.ABI	mid coding region
		LC016.ABI	mid coding region
		LC017.ABI	3' coding region
		LC018.ABI	3' coding region
		LC019.ABI	5' coding region
		LC020.ABI	5' coding region
003	013	AGGTTTGACCCGACTATGCAAGTACAAT	
005	015	TATGCAAGTACAAT	
007	017	CGTTCGCATGCTGTCCATCCTCTTCACT	
009	009	GCATGCTGTCTCCAT	
		AAATGGCTCCAGCACGCCATGGTGCAGT	
		AGCACCGGGCCATGGT	
		CAATGGCAGTGATCAGAAGTCCAAGTGGC	
		GCAGTGATCAGAAGT	
		ATTGCTGCAGCTAAACCATAAAATCTG	
		CTTAAACCATAAAA	
		AAAAAAAGATTCCAAATCTGCATCAGTAAT	
		AAGATTTCACATCTG	
		CAGTGTCTAGGACAGTCACCCCTTTTTC	
		GGACAGTCACCCCTT	
		TGTACCAAATTGCTGTAGACAGTGTACCACT	
		GCTGTTAGACAGTGTAA	
		GTGGAGGGAGCCGTCCCGTACCCCAACC	
		GGACCGTCCCCTTACCC	
		TTAACAGGTGGAATCCATCATGGTGGTGT	
		GGAATCCATCATTGG	

Fig. 6

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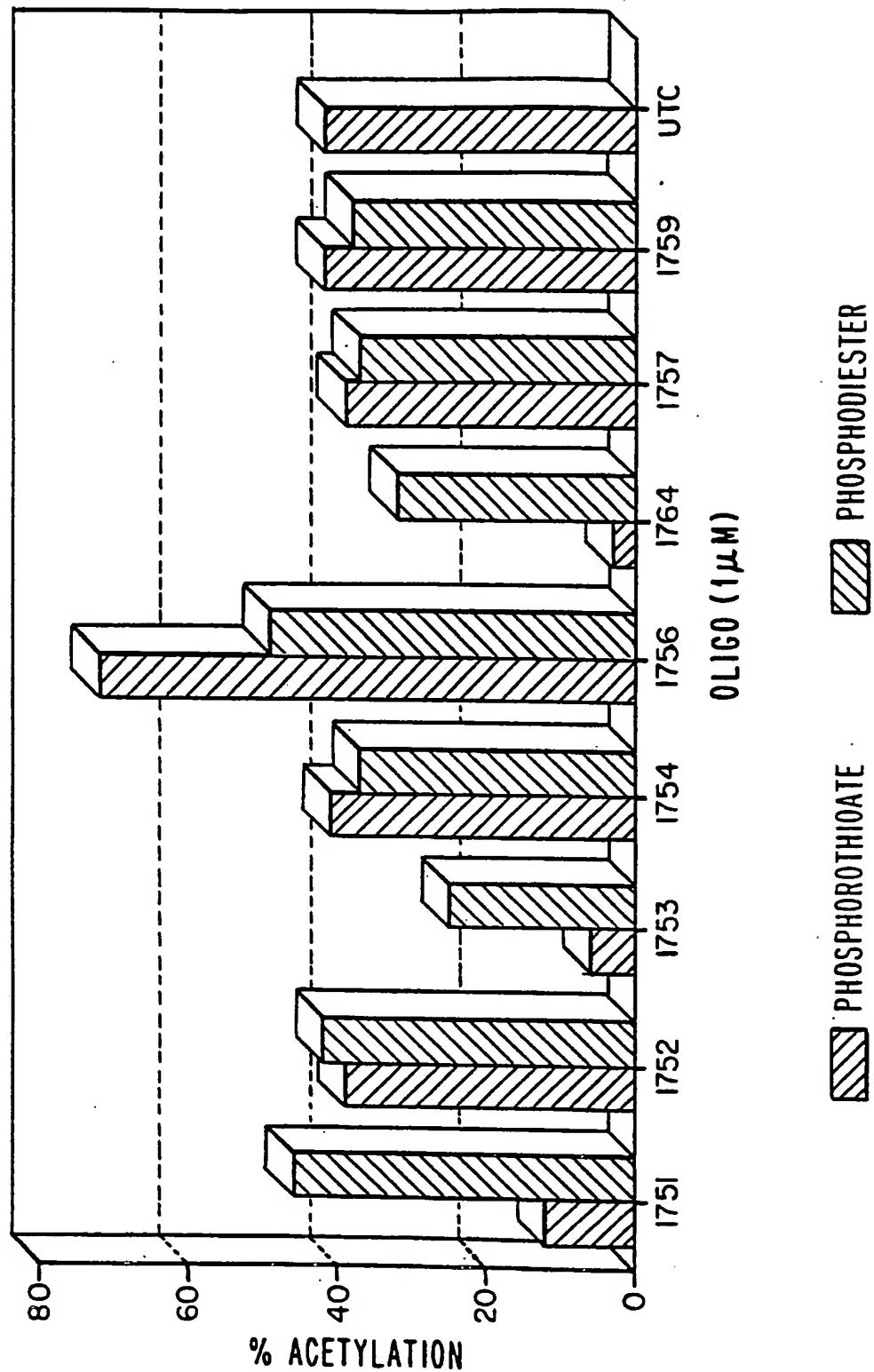
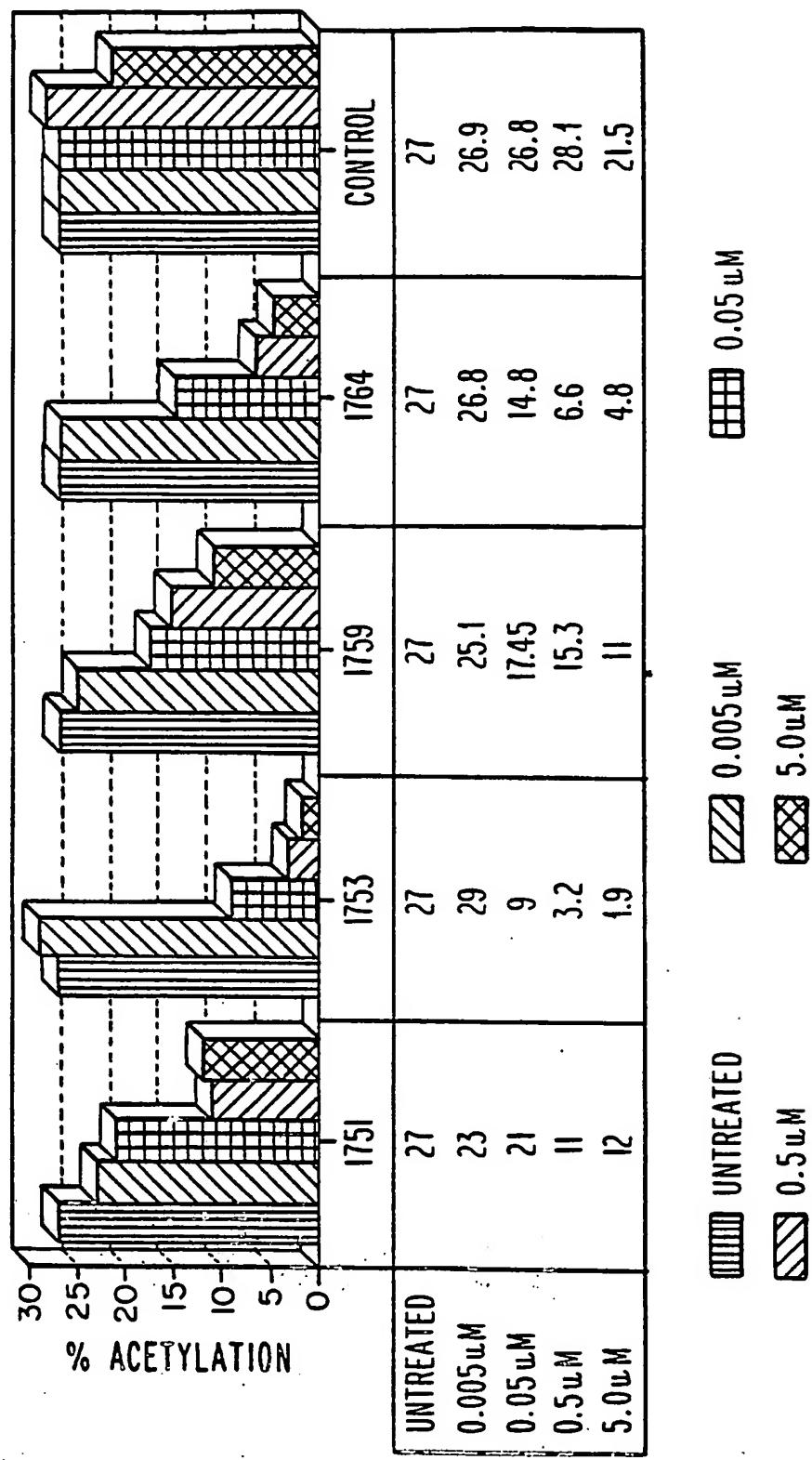


Fig. 7

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*Fig. 8*

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E2 TRANSACTIVATOR AUG REGION

2595	2605	2615	2625
*	*	*	*
AGGAGGAGGAUAGUGAAGAGGAGAACGGCAUGCGAACGUUAACAU			
TCACTTCTCCTTACCTCTGTGCTACGCTTG			
TACCTCTGTGCTACGCC			
TCACTTCTCCTTACCT			
TCACTTCTCCTTACCTCTG			
TCACTTCTCCTTACCTCTGTCG			
TCCTATCACTTCTCC			
TTCTCCTTACCTCTGT			
CCTACCTCTGTGCTGA			
041			

Fig. 9A

E2 TRANSPRESSOR AUG REGION

3 080	3 090	3 100	3 110
*	*	*	*
ACUGGGCUCUACUACUGGCACCAUGGGGGGGUGGUCCGGCAUUUACUA			
TGACGGTGGTACCGGGCCACGGACCTGGCTTAA			
TGTTACGGGCCACGA			
ACGGGGTACGGGCCA			
ATGACGGTACCGGG			
TGATGACGGTGGTACC			
GAGATGATGACGTGG			
045			

Fig. 9B

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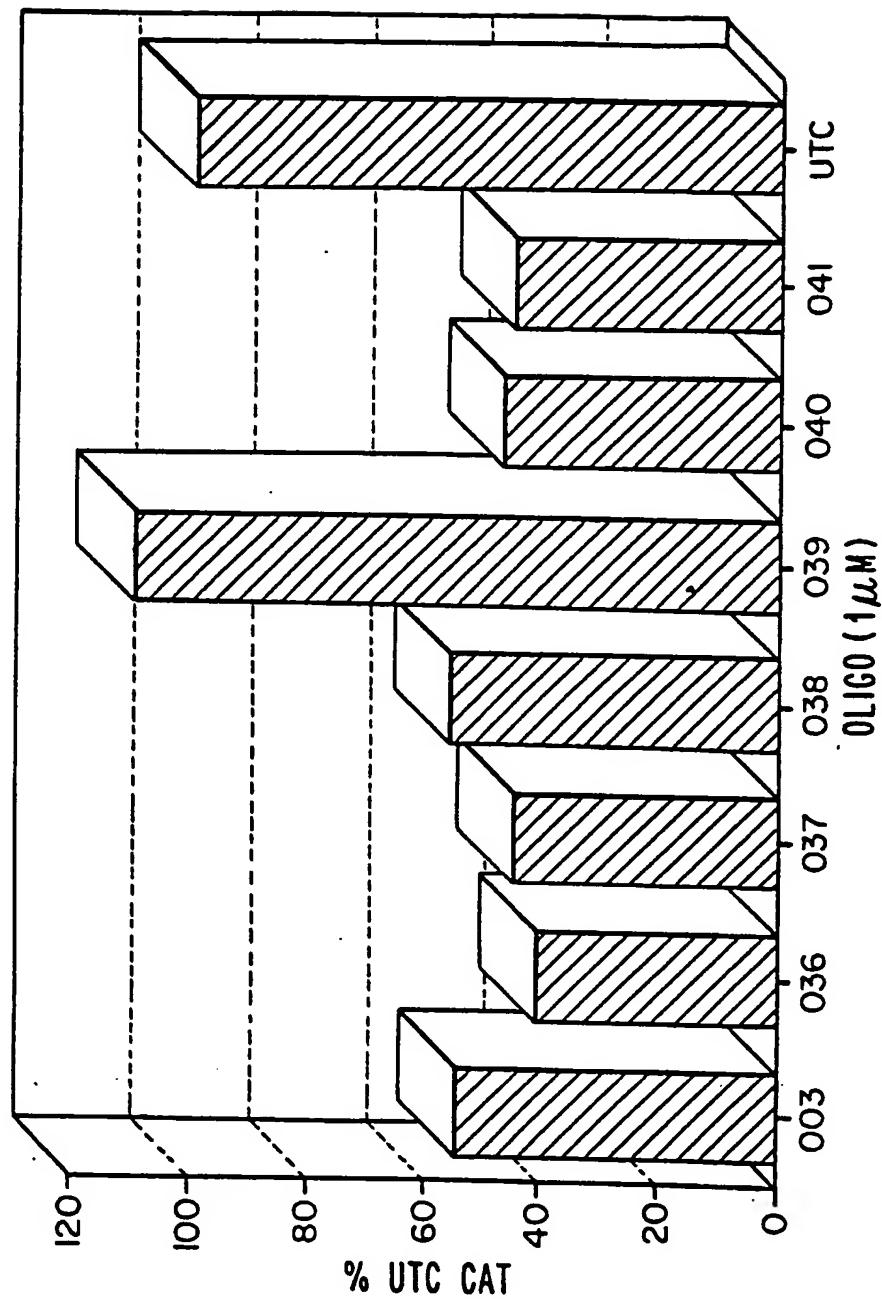


Fig. 10

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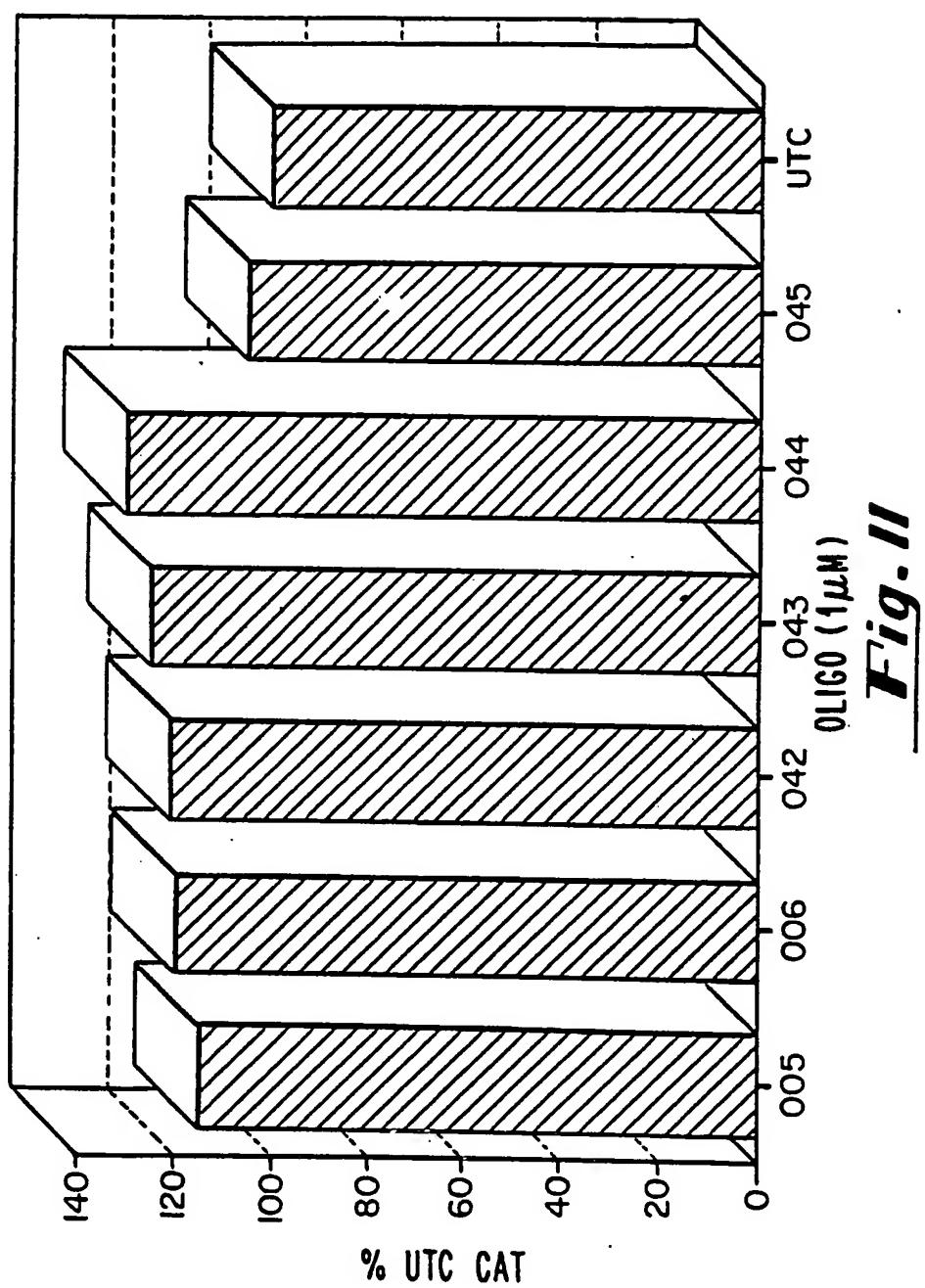
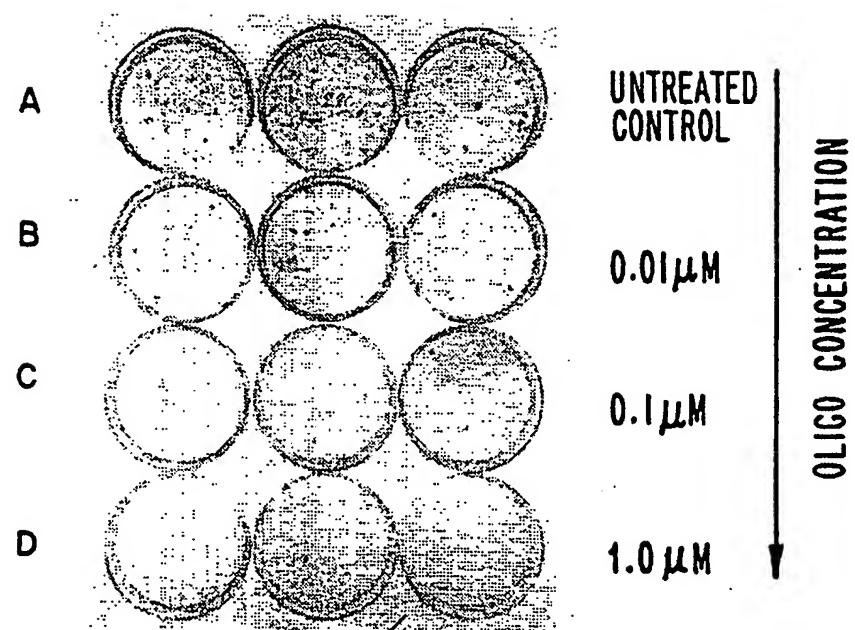


Fig. II

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FIG.12

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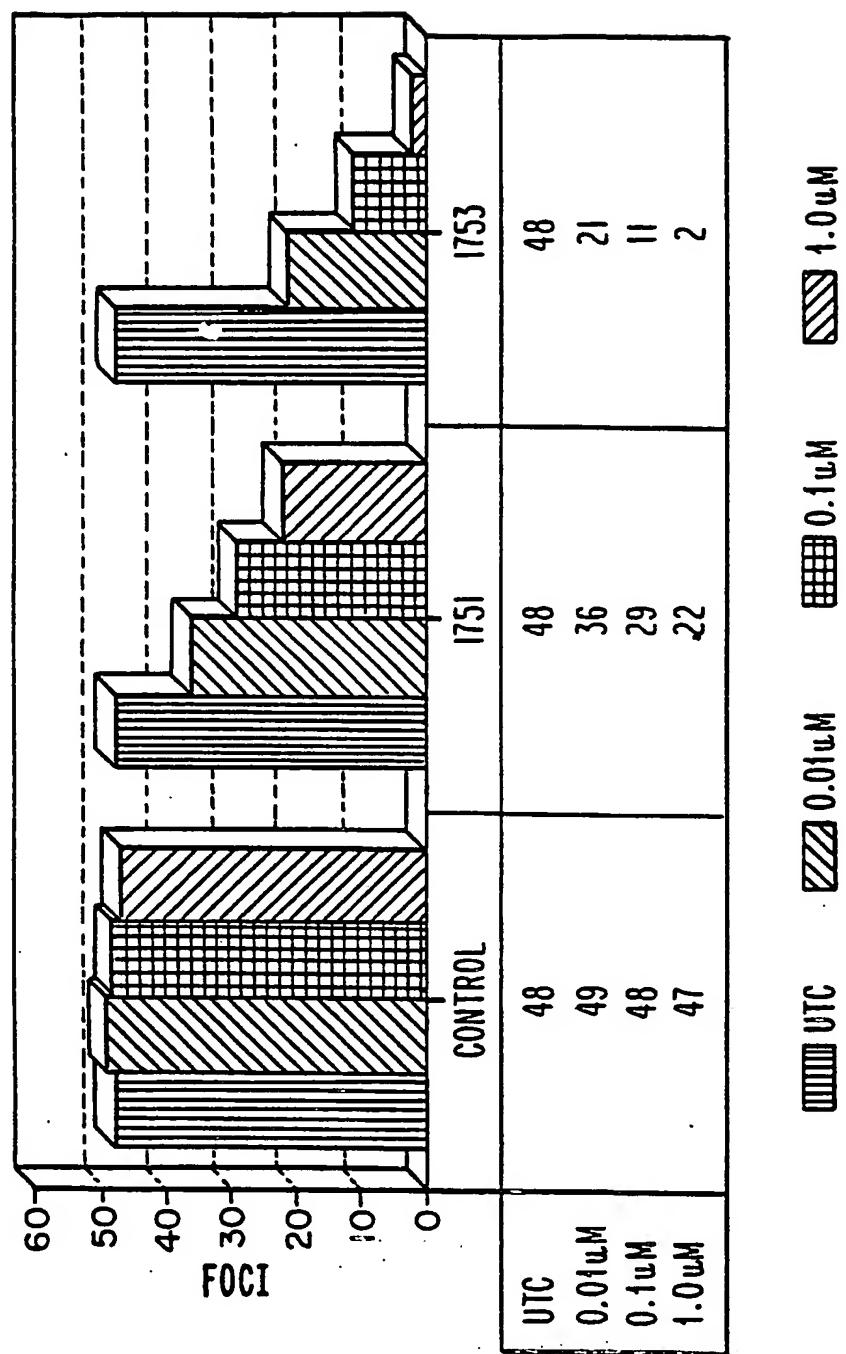


Fig. 13

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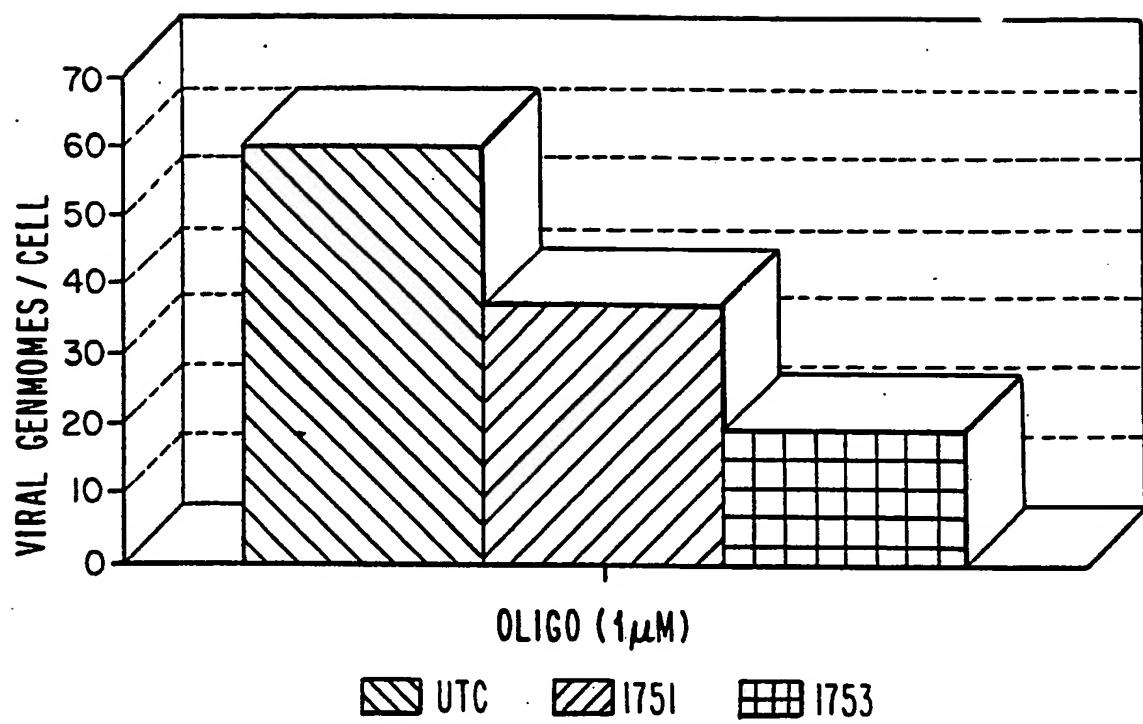
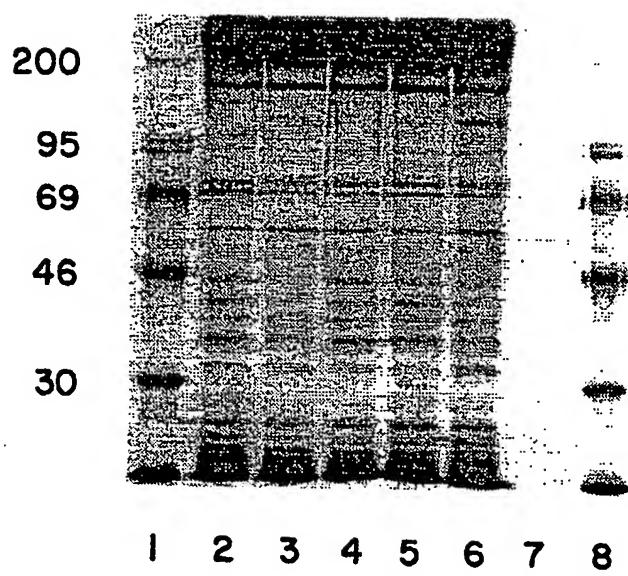


Fig. 14

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FIG. 15

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E2_First_Methionine

2710	2720	2730	2740	2750
*	*	*	*	*
GAT	TCA	GAG	GAC	GAG
CTA	AGT	CTC	CTG	CTC
D	S	E	D	E
<u>C</u>	<u>C</u>	<u>C</u>	<u>C</u>	<u>C</u>
<u>I</u>	<u>Q</u>	<u>R</u>	<u>T</u>	<u>R</u>
<u>d</u>	<u>d</u>	<u>d</u>	<u>d</u>	<u>d</u>

GAT	GGG	AGC	AAT	AGC	CAA	GCG	TTT	AGA	TGC	GTG	C
CTA	CTT	CTA	CTC	CTG	TTA	TCG	TTA	AAA	TCT	ACG	CAC
D	S	E	D	G	S	N	Q	A	F	R	V
<u>C</u>											
<u>I</u>	<u>Q</u>	<u>R</u>	<u>T</u>	<u>K</u>	<u>M</u>	<u>E</u>	<u>A</u>	<u>I</u>	<u>A</u>	<u>K</u>	<u>R</u>
<u>d</u>											

Fig. 16

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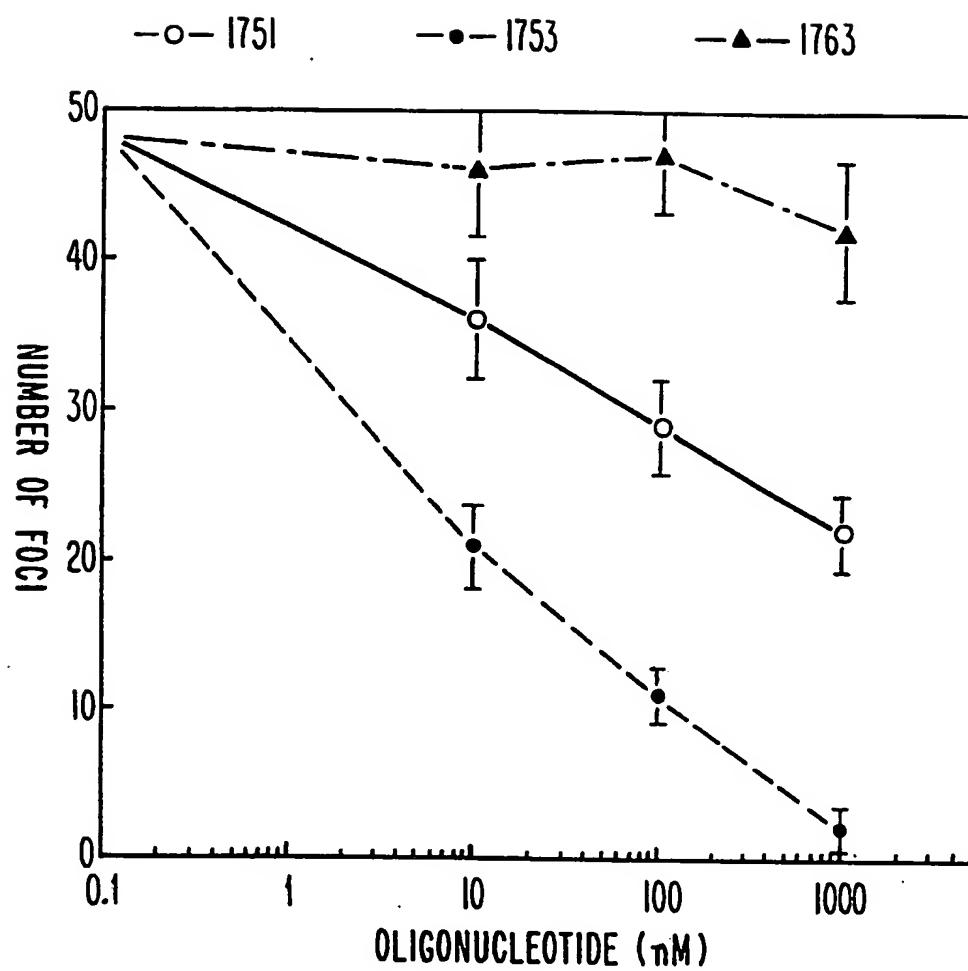


Fig. 17

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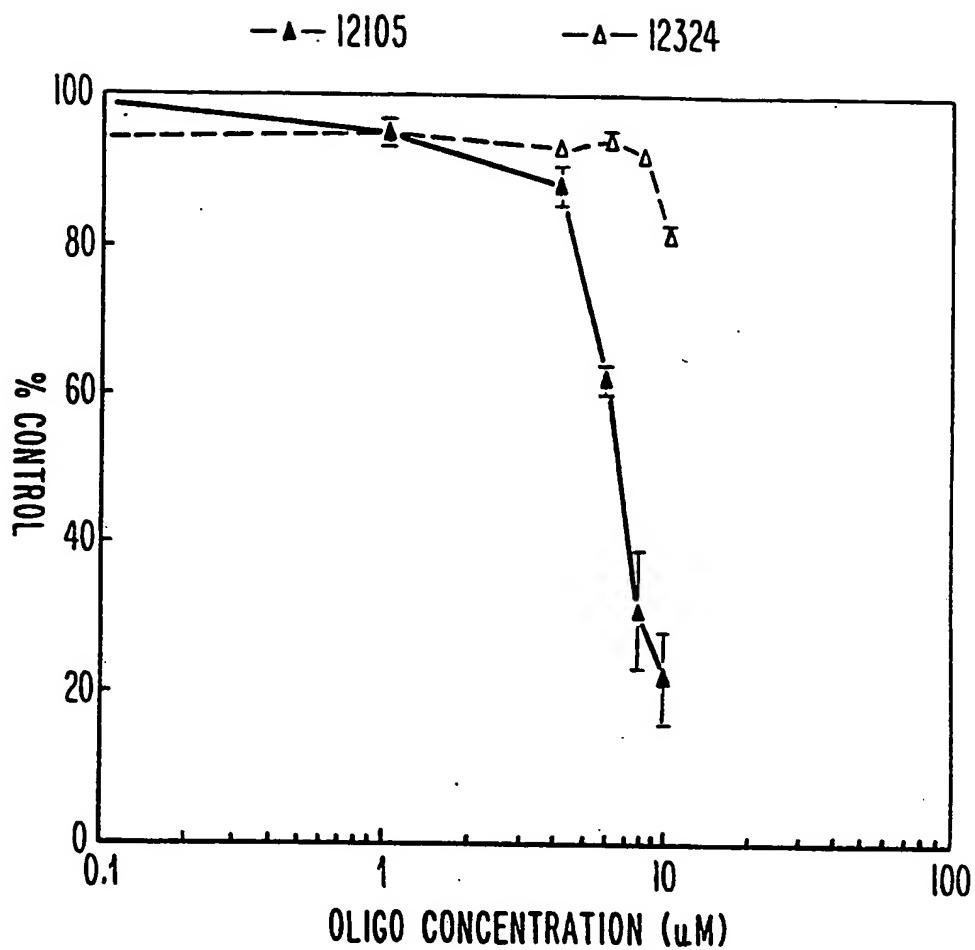


Fig. 18

INTERNATIONAL SEARCH REPORT

International application No.

PCT/US93/03075

A. CLASSIFICATION OF SUBJECT MATTERIPC(S) :C07H 21/04; A61K 48/00
US CL :536/22.1; 514/44

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 536/22.1; 514/44

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

Bios, CABA, Medline, Embase, CA, APS

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	Journal of Virology, Vol. 62, No. 8, issued AUGUST 1988, CHIN, Michael T. et al., "Regulation of Human Papillomavirus Type 11 Enhancer and E6 Promoter by Activating and Repressing Proteins from the E2 Open Reading Frame: Functional and Biochemical Studies", pages 2994-3002, see especially pages 2994, 2996, 2997	1-3
Y	Journal of Molecular Biology, Vol. 193, issued 1987, COLE, S.T. et al., "Nucleotide Sequence and Comparative Analysis of the Human Papillomavirus Type 18 Genome", pages 599-608, see entire document	4-9

<input checked="" type="checkbox"/>	Further documents are listed in the continuation of Box C.	<input type="checkbox"/>	See patent family annex.
"A"	Special categories of cited documents:	"T"	later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
"E"	document defining the general state of the art which is not considered to be part of particular relevance	"X"	document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step
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"P"	document referring to an oral disclosure, use, exhibition or other means		document member of the same patent family

Date of the actual completion of the international search	Date of mailing of the international search report
22 JUNE 1993	30 JUN 1993

Name and mailing address of the ISA/U.S Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231	Authorized officer STEPHANIE W. ZITOMER, PH.D. 
Faxsimile No. NOT APPLICABLE	Telephone No. (703) 308-0196

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US93/03075

C (Continuation). DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	BioTechniques, Vol. 6, No. 10, issued 1988, van der KROL, Alexander R. et al., "Modulation of Eukaryotic Gene Expression by Complementary RNA or DNA Sequences", pages 958-976, see especially page 967 at "Proto-Oncogenes".	7-9